INTERNATIONAL SEARCH REPORT

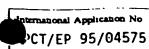
PCT/EP 95/04575

A C! A S	TELECATION OF SUBJECT ASSETS		<u> </u>
ÎPC 6	C12N15/57 C12N9/64 A61K38 C12N5/10 A61K48/00	3/48 G01N33/50	C12Q1/68
According	to International Patent Classification (IPC) or to both national cla	assification and IPC	
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IPC 6	documentation searched (classification system followed by classifi C12N A61K G01N C12Q	, ,	
	tion searched other than minimum documentation to the extent the		
	data base consulted during the international search (name of data	oase and, where practical, search to	erms used)
C. DOCUN	MENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the	e relevant passages	Relevant to claim No.
P,X	CELL, APR 7 1995, 81 (1) P27-40 STATES, XP002010548 RICHARD I ET AL: "Mutations in proteolytic enzyme calpain 3 cal limb-girdle muscular dystrophy see the whole document	the use	1-20
X	JOURNAL OF BIOLOGICAL CHEMISTRY vol. 264, no. 33, 1989, BALTIMOR pages 20106-20111, XP002010549 HIROYUKI SORIMACHI ET AL.: "Mocloning of a novel mammalian calcium-dependent protease distiboth m- and mu-types" cited in the application see figures 2,3	ŔE, MD US, lecular	1-8, 12-15
X Furth	er documents are listed in the continuation of box C.	Patent family members a	are listed in annex.
* Special cate	egories of cited documents :	T later document published after	er the international filing date
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	ctual completion of the international search	Date of mailing of the interna	
	August 1996	2 0. 08. 96	
Name and ma	ailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl, Fax. (+ 31-70) 340-3016	Authonzed officer Espen, J	

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Form PCT/ISA/210 (second sheet) (July 1992)

PC1/EP 95/045/5					
	C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT				
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.			
X	JOURNAL OF BIOLOGICAL CHEMISTRY, vol. 268, no. 26, 1993, BALTIMORE, MD US, pages 19476-19482, XP002010550 HIROYUKI SORIMACHI ET AL.: "A novel tissue-specific calpain species expressed predominantly in the stomach comprises two alternative splicing products with and without Ca2+-binding domain" see figure 2	1-8, 12-15			
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Y	HUMAN MOLECULAR GENETICS, vol. 3, no. 2, February 1994, pages 285-293, XP002010552 FOUGEROUSSE F. ET AL.: "Mapping of a chromosome 15 region involved in limb girdle muscular dystrophy" cited in the application see the whole document	1-20			
Y	JOURNAL OF MEDICAL GENETICS, vol. 30, 1993, pages 385-387, XP000578358 PASSOS-BUENO M. R. ET AL.: "Evidence of genetic heterogeneity in the autosomal recessive adult forms of limb-girdle muscular dystrophy following linkage analysis with 15q probes in Brazilian families" see the whole document	1-20			
Y	GENOMICS, OCT 1994, 23 (3) P619-27, UNITED STATES, XP000578720 RICHARD I ET AL: "Regional localization of human chromosome 15 loci." see page 625	1-20			
A	INT. J. BIOCHEM., vol. 22, no. 8, 1990, pages 811-822, XP002010554 JOHNSON P. ET AL.: "Calpains (intracellular calcium-activated cysteine proteinases): structure-activity relationships and involvement in normal and abnormal cellular metabolism" see the whole document				



C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT Category * Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No.				
Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.			
FASEB JOURNAL, vol. 8, no. 11, 1994,				
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see the whole document				
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	vol. 8, no. 11, 1994, Apages 814-822, XP002010555			

PCT

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

Applicant's or agent's file reference	FOR FURTHER ACTION	See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)
B2628A - FL	International filing date (day/	month/year) Priority date (day/month/year)
International application No.		
PCT/EP 95/ 04575	21/11/1995	22/11/1994
International Patent Classification (IPC) or	`	
	C12N15/57	
Applicant		
ASSOCIATION FRANCAISE CO	NTRE LES MYOPATHIES	et al.
Authority and is transmitted to th 2. This REPORT consists of a tota This report is also accompan	of sheets, including	of the description, claims and/or drawings which have
These annexes consists of a total	ofsheets.	
3. This report contains indications as	nd corresponding pages relating	o the following items:
I X Basis of the report		
II Priority		
	oninion with regard to novelty, i	nventive step and industrial applicability
		•
IV Lack of unity of inven		navelty, inventive step or industrial applicability.
V Reasoned statement un citations and explanati	nder Article 35(2) with regard to ons supporting such statement	novelty, inventive step or industrial applicability;
VI Certain documents cit	ed	
VII Certain defects in the	international application	
	on the international application	
V (2)		
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Date of submission of the demand	Dat	e of completion of this report
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19/06/1996		£ 7. 03. 97
l Name and mailing address of the		norized officer \mathcal{H}
	, P.B. 5818 Patentlaan 2	J. Espen
Tel. (+31-70) 340-2040.	Tx. 31 651 epo ni.	4
Fax: (+31-70) 340-3016	Tele	phone No. +31/70/340-2625
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PCT/EP95/04575

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

I. Basis of the report

1.	This rep invitation amendn	n undel	r Article 14 are re	on the basis of (F eferred to in this r	Replacement sheets report as "originally fil	which have been furi led" and are not anne	nished to the receiving o exed to the report since	Office in respo they do not co	ense to an ontain –
		X	the internationa	d application as or	riginally filed	• -	· ·		-
			the description,	pages			, as originally filed		- -
				pages			, filed with the demand		
				pages			, filed with the letter of		
			-				and also allow Shard		
			the claims, Nos	3.		-	, as originally filed		
			Nos	3.			, as amended under Ar	ticle 19	
			Nos	3 .	•		, filed with the demand		
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		o	the drawings,	sheet͡s / fiḡ.			, as originally filed		
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2	The am	endme	ints have resulte	d in the cancellati	ion of:				
			the description						
		_	the claims, Nos			·.			
		_	the drawings, s						
3.		This o	opinion has beer nd the disclosure	n established as it e as filed (Rule 70	f (some of) the amen 0.2 (c)).	 dments had not beer	n made, since they have	e been conside	ered to go
4.	Addition	nai obs	ervations, if nec	essary:					

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1.	Statement
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Novelty	Claims	9-20	162
	Claims	1-8	NO
Inventive Step	Claims	9-11,15-20	YES
	Claims	1-8,12-14	NO
Industrial Applicability	Claims	1-20	YES
•	Claims		NO

2. Citations and Explanations

1). The present international application relates to Limb-Girdle Muscular Dystrophy Type 2A (LGMD2A). LGMD2A is strongly correlated to mutations in the gene of the proteolytic enzyme calpain 3 (*CANP3*).

The genomic organization of the human *CANP3* gene was determined. The above mentioned enzyme may also be referred to as 'p94' or 'nCL-1'.

2). The following document has been considered for the purposes of this report:

D1 THE JOURNAL OF BIOLOGICAL CHEMISTRY, Vol. 264, pp. 20106-20111,1989

3). The present application does not satisfy the criterion set forth in Article 33(2) PCT because the subject-matter of claims 1-8 is not new in respect of prior art as defined in the regulations (Rule 64(1)-(3) PCT).

Document D1 discloses the nucleotide sequence of the human *CANP3* gene and the deduced amino acid sequence (D1, Fig. 2).

In view of the above comment, D1 is novelty destroying for claims 1-8 (Art. 33 (2) PCT). The involvement of the *CANP3* gene in the etiology of LGMD2 is an implicit feature of the above sequences, and does not render novel the claimed subject-matter.

4). The methods of claims 12-14 are novel (Art. 33 (2) PCT), however they do not involve an inventive step (Art. 33 (3) PCT) for the following reasons.



INTERNATIONAL PRELIMINARY EXAMINATION REPORT

PCT/EP95/04575

For the person skilled in the art, being aware of the known amino acid sequences of claims 5 to 7 or of the known nucleic acid sequences of claims 1-4, the carrying out of said methods required nothing out of the ordinary and thus involved no inventive skill, all being a matter of technical convenience. In addition, this was a matter of normal design procedures for which neither 'creative thinking' nor 'inventive talent' were necessary.

5). Claims 9-11, and 15-20 do meet the requirements of Art. 33 (2) and (3) for the following reasons:

Before the priority date of the present international application, it was neither disclosed nor suggested that the *CANP3* gene product, when mutated, is involved in the LGMD2 disease.

In consequence, the use of sequences related to the *CANP3* gene in the diagnostic of the LGMD2 disease or their use in the preparation of a pharmaceutical composition for the treatment of said disease could not be deduced in an obvious manner from the closest prior art document D1.

The industrial applicability of the present set of claims is acknowledged (Art. 33 (4) PCT).

International application No.

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INTERNATIONAL PRELIMINARY EXAMINATION REPORT

VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:

- -Claim 6 should read '..according to claim 5, ...'.
- -Claim 18 should read '...by nucleic acid amplification...'



PCT

INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference FOR FURTHER see Notification of Transmittal of International Search Report				
B2628A - FL	ACTION (Form PCT/ISA/220) as well as, where applicable, item 5 below.			
International application No.	International filing date(day/month/year)	(Earliest) Priority Date (day/month/year)		
PCT/EP 95/04575	21/11/95	22/11/94		
Applicant				
ASSOCIATION FRANCAISE CON	TRE LES MYOPATHIES et al.			
This international search report has been according to Article 18. A copy is being t	prepared by this International Searching Authoransmitted to the International Bureau.	ority and is transmitted to the applicant		
This international search report consists of X It is also accompanied by a cop	of a total of sheets. y of each prior art document cited in this repor	rt.		
Certain claims were found unsea.	rchable (see Box I).			
2. Unity of invention is lacking (see	Box II).			
3. The international application con international search was carried	ntains disclosure of a nucleotide and/or amino a out on the basis of the sequence listing	acid sequence listing and the		
filed	with the international application.			
X furn	ished by the applicant separately from the inter	rnational application,		
Ĺ	but not accompanied by a statement to the matter going beyond the disclosure in the			
Tran	scribed by this Authority			
4. With regard to the title, the t	ext is approved as submitted by the applicant.			
X the t	ext has been established by this Authority to re	ead as follows:		
LGMD GENE CODING FOR A	CALCIUM DEPENDENT PROTEASE			
6 90		,		
5. With regard to the abstract,	ext is approved as submitted by the applicant			
~ .	ext is approved as submitted by the applicant. ext has been established, according to Rule 38.	2(b), by this Authority as it appears in		
Box	III. The applicant may, within one month fron the report, submit comments to this Authority.			
6. The figure of the drawings to be publis	shed with the abstract is:			
Figure No as su	ggested by the applicant.	X None of the figures.		
=	use the applicant failed to suggest a figure.			
becau	use this figure better characterizes the invention	n.		

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IPC 6	SIFICATION OF SUBJECT MATTER C12N15/57 C12N9/64 A61K38 C12N5/10 A61K48/00	/48 G01N33/50	C12Q1/68
According	to International Patent Classification (IPC) or to both national cla	assification and IPC	
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	tion searched other than minimum documentation to the extent th		
Electronic	data base consulted during the international search (name of data	base and, where practical, search t	erms used)
C. DOCUM	MENTS CONSIDERED TO BE RELEVANT		
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wnich i	nt which may throw doubts on priority claim(s) or s cited to establish the publication date of another or other special reason (as specified)		nen the document is taken alone
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P' docume	neaus ant published prior to the international filing date but an the priority date claimed	in the art.	ing obvious to a person skilled
	actual completion of the international search	& document member of the sar	
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Name and m	ailing address of the ISA	Authorized officer	
	European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl,		
	Fax: (+31-70) 340-2040, 1x. 31 651 epo ni,	Espen, J	

ation) DOCUMENTS CONSIDERED TO BE BELEVANT	PC1/EP 95/045/5
	Relevant to claim No.
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ategory *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.	
	FACED JOHDNAL		
	FASEB JOURNAL,	1	
	vol. 8, no. 11, 1994, pages 814-822, XP002010555		
	pages 814-822, XP002010555		
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WORLD INTELLECTUAL PROPERTY ORGANIZATION International Bureau



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶:
C12N 15/57, 9/64, A61K 38/48, G01N 33/50, C12Q 1/68, C12N 5/10, A61K 48/00

A2

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(21) International Application Number:

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21 November 1995 (21.11.95)

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(81) Designated States: CA, JP, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).

(30) Priority Data:

94402668.1

22 November 1994 (22.11.94) EP

(34) Countries for which the regional or international application was filed:

was filed: GB et al.

(71) Applicant (for all designated States except US): ASSOCIATION FRANÇAISE CONTRE LES MYOPATHIES [FR/FR]; 13, place de Rungis, F-75013 Paris (FR).

(72) Inventors; and

(75) Inventors/Applicants (for US only): BECKMANN, Jacques [FR/FR]; 95, rue de Paris, F-94220 Charenton-le-Pont (FR). RICHARD, Isabelle [FR/FR]; 72, rue de l'Essonne, F-91000 Evry (FR).

(74) Agents: GUTMANN, Ernest et al.; Ernest Gutmann - Yves Plasseraud S.A., 3, rue Chauveau-Lagarde, F-75008 Paris (FR).

(54) Title: LGMD GENE CODING FOR A CALCIUM DEPENDENT PROTEASE

(57) Abstract

A nucleic acid sequence comprising: 1) the sequence represented in figure 8; or 2) the sequence represented in figure 2; or 3) a part of the sequence of figure 2 with the proviso that it is able to code for a protein having a calcium dependant protease activity involved in a LGMD2; or 4) a sequence derived from a sequence defined in 1), 2) or 3) by substitution, deletion or addition of one or more nucleotides with the proviso that said sequences still codes for said protease.

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WORLD INTELLECTUAL PROPERTY ORGANIZATION International Bureau



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 6:
C12N 15/57, 9/64, A61K 38/48, G01N 33/50, C12Q 1/68, C12N 5/10, A61K 48/00

(11) International Publication Number:

WO 96/16175

(43) International Publication Date:

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(21) International Application Number:

PCT/EP95/04575

A2

(22) International Filing Date:

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EP

(71) Applicant (for all designated States except US): ASSOCIATION FRANÇAISE CONTRE LES MYOPATHIES [FR/FR]; 13, place de Rungis, F-75013 Paris (FR).

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(74) Agents: GUTMANN, Ernest et al.; Ernest Gutmann - Yves Plasseraud S.A., 3, rue Chauveau-Lagarde, F-75008 Paris (FR). (81) Designated States: CA, JP, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).

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LGMD gene coding for a calcium dependent protease

The invention relates to the isolated gene coding for a calcium dependent protease belonging to the Calpain family which, when it is mutated, is a cause of a disease called Limb-Girdle Muscular Dystrophy (LGMD).

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The term limb-girdle muscular dystrophy (LGMD) was first proposed by Walton and Nattrass (1954) as part of a classification of muscular dystrophies. LGMD is characterised by progressive symmetrical atrophy and weakness of the proximal limb muscles and by elevated serum creatine kinase. Muscle biopsies demonstrate dystrophic lesions and electromyograms show myopathic features. The symptoms usually begin during the first two decades of life and the disease gradually worsens, often resulting in loss of walking ability 10 or 20 years after onset (Bushby, 1994). Yet, the precise nosological definition of LGMD still remains unclear. Consequently, various neuromuscular diseases such as facioscapulohumeral, Becker muscular dystrophies and especially spinal muscular atrophies have been occasionally classified under this diagnosis. For example, a recent study (Arikawa et al., 1991) reported that 17% (out of 41) of LGMD patients showed a dystrophinopathy. These issues highlight the difficulty in undertaking an analysis of the molecular and genetic defect(s) involved in this pathology.

Attempts to identify the genetic basis of this disease go back over 35 years. Morton and Chung (1959) estimated that "the frequency of heterozygous carrier ... is 16 per thousand persons". The same authors also stated that "the segregation analysis gives no evidence on whether these genes in different families are allelic or at different loci". Both autosomal dominant and recessive transmission have been reported, the latter being more common with an estimated prevalence of 10-5 (Emery, 1991). The localisation of a gene for a recessive form on chromosome 15 (LGMD2A, MIM 253600; Beckmann et al., 1991) provided the definitive proof that LGMD is a specific genetic entity. Subsequent genetic analyses confirmed this chromosome 15 localisation (Young et al., 1992; Passos-Bueno et al., 1993), the latter group demonstrating genetic heterogeneity of this disease. Although a recent study localised a second mutant

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gene to chromosome 2 (LGMD2B, MIM 253601; Bashir et al., 1994), there is evidence that at least one other locus can be involved.

Genetic analyses of the LGMD2 kindreds revealed unexpected findings. First genetic heterogeneity was demonstrated in the highly inbred Indiana Amish community. Second although the Isle of la Réunion families were thought to represent a genetic isolate, at least 6 different disease haplotypes were observed, providing evidence against the hypothesis of a single founder effect (Beckmann et al., 1991) in this inbred population.

The nonspecific nosological definition, the relatively low prevalence and genetic heterogeneity of this disorder limit the number of families which can be used to restrict the genetic boundaries of the LGMD2A interval. Cytogenetic abnormalities, which could have helped to focus on a particular region, have not been reported. Immunogenetic studies of dystrophin-associated proteins (Matsumura et al., 1993) and cytoskeletal or extracellular matrix proteins such as a merosin (Tomé et al., 1994) failed to demonstrate any deficiency. In addition, there is no known specific physiological feature or animal model that could help to identify a candidate gene. Thus, there is no alternative to a positional cloning

It is established that the LGMD2 chromosomal region is localized on chromosome 15 as 15q15.1 - 15q21.1 region (Fougerousse et al., 1994).

Construction and analysis of a 10-12 Mb YAC contig (Fougerousse et al., 1994) permitted the mapping of 33 polymorphic markers within this interval and to further narrow the LGMD2A region to between D15S514 and D15S222. Furthermore, extensive analysis of linkage disequilibrium suggested a likely position for the gene in the proximal part of the contig.

The invention results from the construction of a partial cosmid map and the screening by cDNA selection (Lovett et al., 1991; Tagle et al., 1993) for muscleexpressed sequences encoded by this interval led to the identification of a number of potential candidate genes. One of these, previously cloned by Sorimachi et al. (1989), encodes a muscle specific protein, nCL1 (novel Calpain Large subunit 1), which belongs to the calpain family (CANP, calcium-activated neutral protease; EC 3.4.22.17), and appeared to be a functional candidate gene for this disease.

3 Calpains are non-lysosomal intracellular cysteine proteases which require calcium for their catalytic activities (for a review see Croall D.E. et al, 1991). The mammalian calpains include two ubiquitous proteins CANP1 and CANP2 as well as tissue-specific proteins. In addition to the muscle specific nCL1, stomach specific nCL2 and nCL2' proteins have also been described; these are derived from the same gene by alternative splicing. The ubiquitous enzymes consist of heterodimers with distinct large subunits associated with an common small subunit; the association of tissue-specific large subunits with a small subunit has not yet been demonstrated. The large subunits of calpains can be subdivided into 4 protein domains. Domains I and III, whose functions remain unknown, show no homology with known proteins. Domain I, however, seems important for the regulation of the proteolytic activity. Domain II shows similarity with other cysteine proteases, sharing histidine, cysteine and asparagine residues at its active sites. Domain IV comprises four EF-hand structures which are potential calcium binding sites. In addition, three unique regions with no known homology are present in the muscle-specific nCL1 protein, namely NS, IS1 and IS2, the latter containing a nuclear translocation signal. These regions may be important for the muscle specific function of nCL1.

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It is usually accepted that muscular dystrophies are associated with excess or deregulated calpains, and all the known approaches for curing these diseases are the use of antagonists of these proteases; examples are disclosed in EP 359309 or EP 525420.

The invention results from the finding that, on the opposite to all these hypothesis, the LGMD2 disease is strongly correlated to the defect of a calpain which is expressed in healthy people.

The invention relates to the nucleic acid sequence such as represented in Figure 2 coding for a Ca** dependent protease, or calpain, which is involved in LGMD2 disease, and more precisely LGMD2A. It also relates to a part of this sequence provided it is able to code for a protein having a calcium-dependent protease activity involved in LGMD2, or a sequence derived from one of the above sequences by substitution, deletion or addition of one or more nucleotides provided that said sequence is still coding for said protein, all the nucleic acids yielding a sequence complementary to a sequence as defined above.

The genomic organisation of the human nCL1 gene has been determined by the inventors, and consists of 24 exons and extends over 40 kb as represented in Figure 8, and is also a part of the invention. About 35 kb of this gene have been sequenced. A systematic screening of this gene in LGMD2A families led to the identification of 14 different mutations, establishing that a number of independent mutational events in nCL1 are responsible for LGMD2A. Furthermore, this is the first demonstration of a muscular dystrophy resulting from an enzymatic rather than a structural defect.

In the present specification, CANP3 means the protein which is a Ca** dependent protease, or calpain, and coded by the nCL1 gene on chromosome

The invention relates also to a protein, called CANP3, consisting in the amino acid sequence such as represented in figure 2 and which is involved, when mutated, in the LGMD2 disease.

The cDNA of the gene coding for CANP3, which is coding for the protein, is 15 also represented in Figure 2, and is a part of the invention.

The protein coded by this DNA is CANP3, a calcium-dependent protease belonging to the Calpain family.

Are also included in the present invention the nucleic acid sequences derived from the cDNA of Figure 2 by one or more substitutions, deletions, insertions, or by mutations in 5' or 3' non coding regions or in splice sites, provided that the translated protein has the protease, calcium-dependent activity, and when mutated, induce LGMD2 disease.

The nucleic acid sequence encoding the protein might be DNA or RNA and be complementary to the nucleic and sequence represented in Figure 2. 25

The invention also relates to a recombinant vector including a DNA sequence of the invention, under the control of a promoter allowing the expression of the calpain in an appropriate host cell.

A procaryotic or eucaryotic host cell transformed by or transfected with a DNA sequence comprising all or part of the sequence of Figure 2 is a part of the

Such a host cell might be either:

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- a cell which is able to secrete the protein and, this recombinant protein might be used as a drug to treat the LGMD2, or

- a packaging cell line transfected by a viral or retroviral vector; the cell lines bearing recombinant vector might be used as a drug for gene therapy of LGMD2.

All the systems used today for gene therapy including adenoviruses and retroviruses and others described for example in « l'ADN médicament », (John Libbey, Eurotext, 1993), and bearing one of the DNA sequence of the invention are included herein by reference.

The examples hereunder and attached figures indicate how the structure of 10 the gene was established, and how relationship between the gene and the LGMD was established.

Legend of the figures :

15 Figure 1:

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A) Genomic organisation of the nCL1 gene

The gene covers a 40 kb region of which 35 were sequenced (Accession number pending). Introns and exons are drawn to scale, the latter being indicated by numbered vertical bars. The first intron is the largest one and remains to be fully sequenced. Position of intragenic microsatellites are indicated by asterisks. Arrows indicate the orientation of Alu (closed) and of Mer2 (greyed) repeat sequences.

B) EcoRI restriction map

An EcoRI (E) restriction map of this region was established with the help of cosmids from this region. The location of nCL1 gene is indicated as a black 25 bar. The size of the corresponding fragments are indicated and are underlined when determined by sequence analysis.

C) Cosmid map of the nCL1 gene region.

Cosmids were from a cosmid library constructed by subcloning YAC 774G4 (Richard in preparation) and are presented as lines. Dots on lines 30 indicate positive STSs (indicated in boxed rectangles). A minimum of three cosmids cover the entire gene. T3,T7

6 Figure 2: Sequence of the human nCL1 cDNA (B), and the flanking 5' (A) and 3'

(C) genomic regions.

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A) and C) The polyadenylation signal and putative CAAT, TATAA sites are boxed. Putative Sp1 (position -477 to -472), MEF2 binding sites (-364 to -343) and CArG box (-685 to -672) are in bold. The Alu sequence present in the 5' region is underlined.

B) The corresponding amino acids are shown below the sequence. The coding sequence between the ATG initiation codon and the TGA stop codon is 2466 bp, encoding for a 821 amino acid protein. The adenine in the first methionine codon has been assigned position 1. Locations of introns within the nCL1 gene are indicated by arrowheads. Nucleotides which differ from the previously published ones are indicated by asterisks.

Figure.3: Alignments of amino acid sequences of the muscle-specific calpains.

The human nCL1 protein is shown on the first line. The 3 muscle-specific sequences (NS, IS1 and IS2) are underlined. The second line corresponds to the rat sequence (Accession no P). The third and fourth lines show the deduced amino acid sequences encoded by pig and bovine Expressed Sequences Tagged (GenBank accession no U05678 and no U07858, respectively). The amino acids residues which are conserved among all known members of the calpains are in reverse letters. A period indicates that the same amino acid is present in the sequence. Letters refer to the variant amino acid found in the homologous sequence. Position of missense mutations are given as numbers above the mutated amino acid.

Figure 4: Distribution of the mutations along nCL1 protein structure.

A) Positions of the 23 introns are indicated by vertical bars in relation to the 25 corresponding amino acid coordinates.

B) The nCL1 protein is depicted showing the four domains (I, II, III, IV) and the muscle specific sequences (NS, IS1 and IS2). The position of missense mutations within nCL1 domain are indicated by black dots. The effect of nonsense and frameshift mutations are illustrated as truncated lines, representing the extent of protein synthesised. Name of the corresponding families are indicated on the left of the line. The out of frame ORF is given by hatched lines.

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Figure 5: Northern blot hybridisation of a nCL1 clone

A mRNA blot (Clontech) containing 2 µg of poly(A)+ RNA from each of eight human tissues was hybridised with a nCL1 genomic clone spanning exons 20 and 21. The latter detects a 3.6 kb mRNA present only in a line corresponding to the skeletal muscle mRNA.

Figure 6: Representative mutations identified by heteroduplex analysis.

Examples of mutation screening by heteroduplex analysis. Pedigree B505 shows the segregation of two different mutations in exon 22.

Figure 7: Homozygous mutations in the nCL1 gene

Detection by sequencing of mutations in exons 2 (a), 8 (b), 13 (c) and 22 (d). Sequences from a healthy control are shown above each mutant sequence. Asterisks indicate the position of the mutated nucleotides. The consequences on codon and amino acid residues are indicated on the left of the figure together with the name of the family.

15 Figure 8 : Structure of nCL1 gene

Figure 8A represents the 5' part of the gene with exon 1.

Figure 8B represents the part of the gene including exons 2 to 8,

Figure 8C represents the part of the gene including exon 9,

Figure 8D represents the part of the gene including exons 10 to 24 including the 3' non transcribed region.

EXAMPLES

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EXAMPLE 1

Localisation of the nCL1 within the LGMD2A interval

Detailed genetic and physical maps of the LGMD2A region were constructed (Fougerousse et al., 1994), following the primary linkage assignment to 15q (Beckmann et al., 1991). The disease locus was bracketed between the D15S129 and D15S143 markers, defining the cytogenetic boundaries of the LGMD2A region as 15q15.1-15q21.1 (Fougerousse et al., 1994). Construction and analysis of a 10-12 Mb YAC contig (Fougerousse et al., 1994) permitted us to map 33 polymorphic markers within this interval and to further narrow the LGMD2A region to between D15S514 and D15S222.

The nCL1 gene had been localised to chromosome 15 by hybridisation with sorted chromosomes and by Southern hybridisation to DNA from human-mouse cell hybrids (Ohno et al., 1989).cDNA capture using YACs from the LGMD2A interval allowed the identification of thirteen positional candidate genes. nCL1 was one of the two transcripts identified that showed muscle-specific expression as evidenced by northen blot analysis. The localisation was further confirmed by STS (for Sequence Tagged Site) assays. Primers used for the localisation of the nCL1 gene are P94in2, P94in13 and pcr6a3, as shown in Figure 1 and their characteristics being defined in Table 1.

10 Table 1: PCR primers used for localisation of the nCL1 gene.

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Primer name	Primer sequence (5'-3')	Position within the	Annealing temp (°C)	PCR J	product size on
P94in2	ATCCACCOLLO	cDNA		cDNA	genomic DNA
	ATGGAGCCAACAGAACTGA C GTATGACTCGGAAAAGAAG GT	341-360 428-448	58	108	1758
P94in13	TAAGCAAAAGCAGTCCCCA C TTGCTGTTCCTCACTTTCCT G	1893-1912 1936-1956	58	64	1043
P94-6a3	GTTTCATCTGCTGCTTCGTT CTGGTTCAGGCATACATGG T	2342-2361 2452-2471	56	130	818
P94ex1ter	TTCTTTATGTGGACCCTGAG TT ACGAACTGGATGGGGAACT	218-239 275-293	55	76	76

These primers are designed from different parts of the published human cDNA sequence (Sorimachi et al., 1989), and were used for an STS content screening on DNA from three chromosome 15 somatic cell hybrids and YACs from the LGMD2A contig. The results positioned the gene in a region previously defined as 15q15.1-q21.1 and on 3 YACs (774G4, 926G10, 923G7) localised in this region. The relative positions of STSs along the LGMD2A contig allowed to localise the gene between D15S512 and D15S488, in a candidate region suggested by linkage disequilibrium studies.

The same primers as above were used to screen a cosmid library from YAC 774G4. A group of 5 cosmids was identified (Fig. 1). Experiments with another nCL1 primer pair (P94ex1ter; Table 1) established that these cosmids cover all nCL1 exons except number 1, and that a second group of 4 cosmids contain this

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exon (Fig. 1). A minimal set of three overlapping cosmids (2G8-2B11-1F11) covers the entire gene (Figure 1). DNA from these cosmids was used to construct an *EcoRI* restriction map of this region (Figure 1B).

EXAMPLE 2

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Determination of the nCL1 gene sequence

Most of the sequences were obtained through shotgun sequencing of partial digests of cosmid 1F11 subcloned in M13 and bluescript vectors, and by walking with internal primers. The sequence assembly was made using the XBAP software of the Staden package (Staden) and was in agreement with the restriction map of the cosmids. Sequences of exon 1 and adjacent regions were obtained by sequencing cosmid DNA or PCR products from human genomic DNA. The first intron is still not fully sequenced, but there is evidence that it may be between 10 to 16 kb in length (based on hybridisation of restriction fragments; data not shown). The entire gene, including its 5' and 3' regions, is more than 40 kb long, and shown in Figure 8.

a) the cDNA sequence

The used technology allows the implementation of the published human cDNA sequence of nCL1 (Sorimachi 1989). It contains the missing 129 bases corresponding to the N-terminal 43 amino acids (Figure 2). It also differs from it at 12 positions. Three of which occur at third base positions of codons and preserve the encoded amino acid sequence. The other 9 differences lead to changes in amino-acid composition (Figure 2). As these different exons were sequenced repeatedly on at least 10 distinct genomes, we are confident that the sequence of Fig. 2 represents an authentic sequence and does not contain minor polymorphic variants. Furthermore, these modifications increase the local similarity with the rat nCL1 amino acid sequence (Sorimachi), although the overall similarity is still 94 %.

The ATG numbered 1 in Figure 2 is the translation initiation site based on homology with the rat nCL1, and is within a sequence with only 5 nucleotides out of 8 in common with the Kosak consensus sequence (Kosak M, 1984). Putative CCAAT and TATA boxes were observed 590, 324, (CCAAT) and 544 or 33 bp (TATA) upstream of the initiating ATG codon, respectively (Bucher, 1990). A GC-box binding the Sp1 protein (Dynan et al., 1983) was identified at position -477.

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Consensus sequences corresponding to potential muscle-specific regulatory elements were identified (Fig. 2). These include a myocyte-specific enhancer-binding factor 2 (MEF2) binding site (Cserjesi P. 1991), a CArG box (Minty A. 1986) and 6 E-boxes (binding sites for basic Helix-Loop-Helix proteins frequently found in members of MyoD family; Blackwell et Weintraub, 1990). The functional significance of these putative transcription factor binding sites in the regulation of nCL1 gene expression remains to be established.

Two potential AAUAAA polyadenylation signals, were identified 520 and 777 bp downstream of the TGA stop codon. The sequencing of a partial nCL1 cDNA containing a polyA tail, demonstrated that the first AAUAAA is the polyadenylation signal. The latter is embedded in a region well conserved with the rat nCL1 sequence and is followed after 4 bp by a G/T cluster, present in most genes 3' of the polyadenylation site (Birnstiel et al., 1985). The 3'-untranslated region of the nCL1 mRNA is 565 bp long. The predicted length of the cDNA should therefore be approximately 3550 or 3000 bp.

b) Comparison with calpain

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The sequence of the human nCL1 gene was compared to those of other calpains thereof (Figure 3). The most telling comparisons are with the homologous rat (Accession no J05121), bovine (Accession no U07858) and porcine (Accession no U05678) sequences. The accession numbers refers to those or international genebanks, such as GeneBank (N.I.H.) or EMBL Database (EMBL, Heidelberg). High local similarities between the human and rat DNA sequences are even observed in the 5' (75%) or in different parts of the 3' untranslated regions (over 60%) (data not shown). The high extent of sequence homology manifested by the human and rat nCL1 gene in their untranslated regions is suggestive of evolutionary pressures on common putative regulatory sequences.

c) Genomic organisation of the nCL1 gene

A comparison of the published nCL1 human cDNA (Sorimachi et al., 1989) with the corresponding genomic sequence led to the identification of 24 exons ranging in length from 12 bp (exon 13) to 309 bp (exon 1), with a mean size of 100 bp (Figure 1). The size of introns ranges from 86 bp to about 10-16 kb for intron 1.

The intron-exon boundaries as shown in Table 2 exhibit close adherence to 5' and 3' splice site consensus sequences (Shapiro and Senapathy, 1987).

<u>Table 2:</u> Sequences at the intron-exon junctions. A score expressing adherence to the consensus was calculated for each site according to Shapiro and Senapathy (1987). Sequences of exons and introns are in upper and lower cases, respectively. Size of exons are given in parenthesis.

splice donor site	scoi (%)		score (%)	splice acceptor site Exon	
				Exon 1 (309 bp) ->
CTCCGgtgagt	8 8.5	<-Intron 1->	99.0	tttttgtttcacagGAAAT Exon 2 (70 bp	
GCTAGgtagga	. 83.5	<-Intron 2->	90.0	gtgtctgcctgcagGGGAC Exon 3 (119 bp)	
TCCAGgtgagg	92	<-Intron 3->	81.5	acgcttctgtgcagTTCTG Exon 4 (134 bp)	
GCTAAgtaagc	82	<-Intron 4->	81.5	atcctctctctaagGCTCC Exon 5 (169 bp)	
TTGATgtaagt	.87	<-Intron 5->	79.5	ccatcgggcctcagGATGG Exon 6 (144 bp)	
CCCGGgggg	77.5	<-Intron 6->	91	Hantana - A C A A T	
ATGAGgtaagc	94	<-intron 7->	78.5	(0.00)	
GATAGgtaggt	89	<-Intron 8->	91.5	(00 op) -	
TTCTGgtgagt	88	<-Intron 9->	92	(· · · · · · · · · · · · · · · · · · ·	
.CCCAGgtggga	80	<-Intron 10->	68.5	ticcaacctctcagGATGT Exon 10 (161 bp) =:tictgggggtgcagATACT Exon 11 (170 bp) =:	
.ACGAGgtgtgt	85.5	<-intron 11->	8 6	-	
.AAGAGgtatag	70	<-Intron 12->	87		
TCTGAgtgagt	76.5	<-Intron 13->	97	tccccatctctcagATGCA Exon 13 (209 bp) ->	
CAGTGgtgagt	8 9	<-Intron 14->	93.5	tgtattcctcacagGGAAG Exon 14 (37 bp) ->	
CCAAGgtaggt	89	<-Intron 15->	87	ctttcttatgcagAAAAA Exon 15 (18 bp) ->	
CACAGgigici	80	<-Intron 16->	88	cctcctctctccagCCCAT Exon16 (114 bp) ->	
GAGATgigagi	84	<-Intron 17->	92.5	ttgtgcctccacagCCACA Exon 17 (78 bp) ->	
CAAACgtgagt	83	<-Intron 18->		cccttcctcctcagGACAT Exon 18 (58 bp) ->	
FGGAT gtatcc	56	<-Intron 19->		ciccatcccccagACAAG Exon 19 (65 bp) ->	
GGCAGgtggga	8 0	<-Intron 20->		cctccctcctccagACAGA Exon 20 (69 bp) ->	
GCAGgigcig	66	<-intron 21->		ttttctattgccagAAATA Exon 21 (79 bp) ->ggtcccctccacagGATTC Exon 22 (117 bp) ->	

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GTTCAgtaagt	79	<-Intron 22->	93.5	gcattctttcacagGAGCT	Exon 23	(59 bp) ->
TGGAGgtaaag	81	<-Intron 23->	79	gggacttctttcagTGGCT	Exon 24	(27 bp) ->

When the genomic sequence was submitted to GRAIL analysis (Uberbacher et al., 1991), 11 exons were correctly recognised, 4 were not identified, 6 were inadequately defined and 2 were too small to be recognised (data not shown).

As already noted, the nCL1 gene has three unique sequence blocks, NS (amino acid residues 1 to 61), IS1 (residues 267 to 329) and IS2 (residues 578 to 653). It is interesting to note that each of these sequences, as well as the nuclear translocation signal inside IS2, are essentially flanked by introns (Fig. 4). The exon-intron organisation of the human nCL1 is similar to that reported for the chicken CANP (the only other large subunit calpain gene whose genomic structure is known; (Emori et al., 1986).

Four microsatellite sequences were identified. Two of them are in the distal part of the first intron: an (AT)14 and an previously identified mixed-pattern microsatellite, S774G4B8, which was demonstrated to be non polymorphic (Fougerousse et al., 1994). A (TA)7(CA)4(GA)13 was identified in the second intron and genotyping of 64 CEPH unrelated individuals revealed two alleles (with frequencies of 0.10 and 0.90). The fourth microsatellite is a mixed (CA)n(TA)m repeat present in the 9th intron. The latter and the (AT)14 repeat have not been investigated for polymorphism. Fourteen repetitive sequences of the Alu family and one Mer2 repeat were identified in the nCL1 gene (Fig. 1C), which has, thus, on the average one Alu element per 2.5 kb.

Southern blot experiments (Ohno et al., 1989) and STS screening (data not shown) suggest that there is but one copy per genome of this member of the calpain family.

EXAMPLE 3

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Expression of the nCL1 gene

The pattern of tissue-specificity was investigated by northern blot hybridisation with a genomic subclone probe from cosmid 1F11 spanning exons 20 and 21. There is no evidence for the existence of an alternatively spliced form of nCL1, although this cannot be excluded. A transcript of about 3.4-3.6 kb was

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detected in skeletal muscle mRNA (Figure 5). This size therefore favours that the position -544 is the functional TATA box.

Transcription studies suggested that it is an active gene rather than a pseudogene and its muscle-specific pattern of expression is consistent with the phenotype of this disorder (Sorimachi et al., 1989 and Figure 5).

EXAMPLE 4

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Mutation screening

nCL1 fulfils both positional and functional criteria to be a candidate gene for LGMD2A. To evaluate its role in the etiology of this disorder, nCL1 was systematically screened in 38 LGMD2 families for the presence of nucleotide changes using a combination of heteroduplex (Keen et al., 1991) and direct sequence analyses.

PCR primers were designed to specifically amplify the exons and splice junctions and also the regions containing the putative CAT, TATA boxes and the polyadenylation signal of the gene as shown in Table 3.

Table 3: PCR primers used for the analysis of the nCL1 gene in LGMD patients.

amplified region promotor	Primer sequences (5'-3')	Size (bp)	Annealing temp. (°C)
F	TTCAGTACCTCCCGTTCACC	296	59
exon 1	GATGCTTGAGCCAGGAAAAC		•
CAON 1	CTTTCCTTGAAGGTAGCTGTAT	438	60
exon 2	GAGGTGCTGAGTGAGAGGAC		00
c.ton 2	ACTCCGTCTCAAAAAAATACCT	239	57
exon 3	ATTGTCCCTTTACCTCCTGG		27
CAUII 3	TGGAAGTAGGAGAGTGGGCA	354	58
au 1	GGGTAGATGGGTGGGAAGTT	3.74	38
exon 4	GAGGAATGTGGAGGAAGGAC	292	50
	TTCCTGTGAGTGAGGTCTCG	272	59
exon 5	GGAACTCTGTGACCCCAAAT	325	• •
_	TCCTCAAACAAACATTCGC	323	5 6
exon 6	GTTCCCTACATTCTCCATCG	315	
	GTTATTTCAACCCAGACCCTT	313	57
exon 7	AATGGGTTCTCTGGTTACTGC	222	
	AGCACGAAAAGCAAAGATAAA	333	56
exon 8	GTAAGAGATTTGCCCCCCAG	201	
	TCTGCGGATCATTGGTTTTG	321	58
exon 9	CCTTCCCTTCTTCCTGCTTC		
	CTCTCTTCCCCACCCTTACC	173	56
exon 10	CCTCCTCACCTGCTCCCATA		
	TTTTTCGGCTTAGACCCTCC	251	56
exon 11	TGTGGGGAATAGAATAAATGG		
	CCAGGAGCTCTGTGGGTCA	355	57
exon 12	GGCTCCTCATCCTCATTCACA		
	GTGGAGGAGGGTGAGTGTGC	312	61
exon 13	TGTGGCAGGACAGGACGTTC		
	COCAGGACAGGACGTTC	337 ·	60

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	14			
	TTCAACCTCTGGAGTGGGCC			
exon 14	CACCAGAGCAAACCGTCCAC	230	61	
	ACAGCCCAGACTCCCATTCC		0,	
exon 15	TTCTCTTCTCCCTTCACCCT	225	57	
	ACACACTTCATGCTCTCTACCC		,	
exon 16	CCGCCTATTCCTTTCCTCTT	331	56	
	GACAAACTCCTGGGAAGCCT	55.	٠٠.	
exon 17	ACCTCTGACCCCTGTGAACC	270	61	
	TGTGGATTTGTGTGCTACGC	2.0	01	
exon 18	CATAAATAGCACCGACAGGGA	258	59	
	GGGATGGAGAAGAGTGAGGA	200	29	
exon 19	TCCTCACTCTTCTCCATCCC	159	57	
	ACCCTGTATGTTGCCTTGG	137	37	
exons 20-21	GGGGATTTTGCTGTGTGCTG	333	<i>(</i> 1	
	ATTCCTGCTCCCACCGTCTC	333	61	
exon 22	CACAGAGTGTCCGAGAGGCA	282	67	
	GGAGATTATCAGGTGAGATGCC	202	57	
exons 22-23	CAGAGTGTCCGAGAGGCAGGG	608	61	
	CGTTGACCCCTCCACCTTGA	000	61	
exon 24	GGGAAAACATGCACCTTCTT	375	**	
	TAGGGGGTAAAATGGAGGAG	313	58	
polyadenylation signal	ACTAACTCAGTGGAATAGGG	413		
	GGAGCTAGGATAGCTCAAT	413	56	

PCR products made on DNA from blood of specific LGMD2A patients were then subjected either to heteroduplex analysis or to direct sequencing, depending on whether the mutation, based on haplotype analysis, was expected to be homozygous or heterozygous, respectively. It was occasionally necessary to clone the PCR products to precisely identify the mutations (i.e., for microdeletions or insertions and for some heterozygotes). Disease-associated mutations are summarised in Table 4 hereunder and their position along the protein is shown in Fig. 4.

Table 4: nCL1 mutations in LGMD2A families.

Codons and amino acid positions are numbered on the basis of the cDNA sequence starting from ATG.

Exon	Families	Nucleotide position	Nucleotide change	Amino acid	Amino acid	Restriction si
2	B519*	328	CGA->TGA	110	Arg->stop	
4	M42	545	C <u>T</u> G -> C <u>A</u> G	182	Leu->Gin	
4	M1394: M2888	550	CAA -> CA	184	frameshift	
5	M35: M37	701	GGG -> GAG	234	Gly->Glu	

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6	M32	0.15	15			
Ü	14137	945	CGG -> CG	315	frameshift	-Smal
8	M2407*	1061	G <u>T</u> G -> G <u>G</u> G	354	Val-> Gly	
8	M1394	1079	T <u>G</u> G -> T <u>A</u> G	360	Trp->stop	-Bstnl, -Eco
11	M2888	1468	CGG -> TGG	490	Arg->Trp	
13	R12*	1715	C <u>G</u> G -> C <u>A</u> G	572	Arg->Gin	-Mspl
19	R27	2069-2070	deletion AC	690	frameshift	
21	R14: R17	2230	<u>A</u> GC -> <u>G</u> GC	744	Ser->Gly	-Alul
22	A*: B501*: M32	2306	C <u>G</u> G -> C <u>A</u> G	769	Arg->Gin	
22	B505	2313-2316	deletion AGAC	771-772	frameshift	
	R14: B505	2362-2363	AG -> TCATCT	788	frameshift	

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The first letter of the family code refers to the origin of the population B= Brazil, M= metropolitan France, R = Isle of La Réunion, A= Amish.

Each mutation was confirmed by heteroduplex analysis, by sequencing of both strands in several members of the family or by enzymatic digestion when the mutation resulted in the modification of a restriction site. Segregation analyses of the mutations, performed on DNAs from all available members of the families, confirmed that these sequence variations are on the parental chromosome carrying the LGMD2A mutation. To exclude the possibility that the missense substitutions might be polymorphisms, their presence was systematically tested in a control population: none of these mutations was seen among 120 control chromosomes from the CEPH reference families.

EXAMPLE 5:

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Analysis of families genes, chromosome-15 ascertained families

The initial screening for causative mutations was performed on families, each containing a LGMD gene located on chromosome 15. These included families from the Island of La Réunion (Beckmann et al., 1991), from the Old Order Amish from northern Indiana (Young et al., 1992,) and 2 Brazilian families (Passos Bueno et al., 1993).

a) Reunion Island families

Genealogical studies and geographic isolation of the families from the Isle of La Réunion were suggestive of a single founder effect. Genetic analyses are,

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however, inconsistent with this hypothesis as the families present haplotype heterogeneity. At least, six different carrier chromosomes are encountered, (with affected individuals in several families being compound heterozygotes). Distinct mutations corresponding to four of these six haplotypes have been identified thus far.

In family R14, exons 13, 21 and 22 showed evidence for sequence variation upon heteroduplex analysis (Fig. 6). Sequencing of the associated PCR products revealed (i) a polymorphism in exon 13, (ii) a missense mutation (A->G) in exon 21 transforming the Ser⁷⁴⁴ residue to a glycine in the loop of the second EF-hand in domain IV of the protein (Figure 4), and (iii) a frameshift mutation in exon 22. The exon 21 mutation and the polymorphism in exon 13 form an haplotype which is also encountered in family R17. Subcloning of the PCR products was necessary to identify the exon 22 mutation. Sequencing of several clones revealed a replacement of AG by TCATCT (data not shown). This frameshift mutation causes premature termination at nucleotide 2400 where an in frame stop codon occurs (Figure. 4).

The affected individuals in family R12 are homozygous for all markers of the LGMD2A interval (Allamand, submitted). Sequencing of the PCR products of exon 13 revealed a G to A transition at base 1715 of the cDNA resulting in a substitution of glutamine for Arg⁵⁷² (Figure. 7) within domain III, a residue which is highly conserved throughout all known calpains. This mutation, detectable by loss of *Mspl* restriction site, is present only in this family and in no other examined LGMD2A families or unrelated controls.

In family R27, heteroduplex analysis followed by sequencing of the PCR products of an affected child revealed a two base pair deletion in exon 19 (Figure. 6 and table 4). One AC out of three is missing at this position of the sequence, producing a stop codon at position 2069 of the cDNA sequence (Figure 4).

b) Amish families

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As expected, due to multiple consanguineous links, the examined LGMD2A Northern Indiana Amish patients were homozygous for the haplotype on the chromosome bearing the mutant allele (Allamand, submitted). A (G->A) missense mutation was identified at nucleotide 2306 within exon 22 (Fig. 7). The

resulting codon change is CGG to CAG, transforming Arg⁷⁶⁹ to glutamine. This residue, which is conserved throughout all members of the calpain family in all species, is located in domain IV of the protein within the 3rd EF-hand at the helix-loop junction (ref). This mutation was encountered in a homozygous state in all patients from 12 chromosome 15-linked Amish families, in agreement with the haplotype analysis. We also screened six Southern Indiana Amish LGMD families, for which the chromosome 15 locus was excluded by linkage analyses (Allamand ESHG, submitted, ASHG 94). As expected, this nucleotide change was not present in any of the patients from these families, thus confirming the genetic heterogeneity of this disease in this genetically related isolate.

c) Brazilian families

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As a result of consanguineous marriages, two Brazilian families (B501, B519) are homozygous for extended LGMD2A carrier haplotypes (data not shown). Sequencing PCR products from affected individuals of these families demonstrated that family B501 has the same exon 22 mutation found in northern Indiana Amish patients (Figure 7), but embedded in a completely different haplotype. In family B519, the patients carry a C to T transition in exon 2, replacing Arg³²⁸ with a TGA stop codon (Figure 7), thus leading, presumably, to a very truncated protein (Figure 4).

d) Analysis of other LGMD families

Having validated the role of the candidate gene in the chromosome 15 ascertained families, we next examined by heteroduplex analysis LGMD families for which linkage data were not informative. These included one Brazilian (B505) and 13 metropolitan French pedigrees.

Heteroduplex bands were revealed for exons 1, 3, 4, 5, 6, 8, 11, 22 of one or more patients (Figure 6). Of all sequence variants, 10 were identified as possible pathogenic mutations (5 missense, 1 nonsense and 4 frameshift mutations) and 3 as polymorphisms with no change of amino acid of the protein. All causative mutations identified are listed in Table 4 here-above. Identical mutations were uncovered in apparently unrelated families. The mutations shared by families M35 and M37, and M2888 and M1394, respectively, are likely to be the consequence of independent events since they are embedded in different marker haplotypes. In contrast, it is likely that the point mutation in exon

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22 of the Amish and in the M32 kindreds corresponds to the same mutational event as both chromosomes share a common four marker haplotype (774G4A1-774G4A10-774G454D-774G4A2) around nCL1 (data not shown), possibly reflecting a common ancestor. The same holds true for the AG to TCATCT substitution mutation encountered in exon 22 in families B505 and R14. The exon 8 (T->G) transversion is present in the two carrier chromosomes of M2407, the only metropolitan family homozygous by haplotype, possibly reflecting an undocumented consanguinity. For some families, no disease-causing mutation has been detected thus far (M40 for example).

In addition to the polymorphism present in exon 13 in families R14 and R17 (position 668) and in the intragenic microsatellites, four additional neutral variations were detected: a (T->C) transition at position 96, abolishing a *Ddel* restriction site in exon 1 in M31; a (C->T) transition in exon 3 (position 495) in M40 and in M37 forming a haplotype with the exon 5 mutation (in the former family, this polymorphism does not cosegregate with the disease); a (T->C) transition in the paternally derived promotor in M42 at position -428, which was also evidenced in healthy controls; and a variable poly(G) in intron 22 close to the splice site in families R20, R11, R19, M35 and M37. The latter is also present in the members of the CEPH families, but is not useful as a genetic marker as the visualisation and interpretation of mononucleotide repeat alleles is difficult.

In total, sixteen independent mutational events representing fourteen different mutations were identified. All mutations cosegregate with the disease in LGMD2A families. The characterised morbid calpain alleles contain nucleotide changes which were not found in alleles from normal individual. The discovery of two nonsense and five frameshift mutations in nCL1 supports the hypothesis that a deficiency of this product causes LGMD2A. All seven mutations result in a premature in-frame stop codon, leading to the production of truncated and presumably inactive proteins (Figure 4). Evidences for the morbidity of the missense mutations come from (1) the relative high incidence of such mutations among LGMD2A patients; although it is difficult in the absence of functional assays to differentiate between a polymorphism and a morbid mutation, the occurrence of different "missense" mutations in this gene cannot all be

accounted for as rare private polymorphisms; (2) the failure to observe these mutations in control chromosomes; and (3) the occurrence of mutations in evolutionarily conserved residues and/or in regions of documented functional importance. Four of seven missense mutations change an amino acid which is conserved in all known members of the calpain family in all species (Figure 3). Two of the remaining mutations affect less conserved amino acid residues, but are located in important functional domains. The substitution V354G in exon 8 is 4 residues before the asparagine at the active site and S744G in exon 21 is within the loop of the second EF-hand and may impair the calcium-dependent regulation of calpain activity or the interaction with a small subunit (Figure 4). Several missense mutations change a hydrophobic residue to a polar one, or vice versa (Table 4) possibly disrupting higher order structures.

METHODS

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Description of the patients

The LGMD2A families analysed were from 4 different geographic origins. They included 3 Brazilian families, 13 interrelated nuclear families from the Isle of la Réunion, 10 French metropolitan families and 12 US Amish families. The majority of these families were previously ascertained to belong to the chromosome 15 group by linkage analysis (Beckmann, 1991; Young, Passos-Bueno et al., 1993). However, some families from metropolitan France as well as one Brazilian family, B505, had non significant lodscores for chromosome 15. Genomic DNA was obtained from peripheral blood lymphocytes.

Sequencing of cosmid c774G4-1F11 and EcoRI restriction map of cosmids.

Cosmid 1F11 (Figure 1C) was subcloned following DNA preparation through Qiagen procedure (Qiagen Inc., USA) and partial digestion with either Sau3A, Rsal or Alul. Size-selected restriction fragments were recovered fom low-melting agarose and eventually ligated with M13 or Bluescript (Stratagene, USA) vectors. After electroporation in E.coli, recombinant colonies were picked in 100 µl of LB/ampicillin media. PCR reactions were performed on 1 µl of the culture in 10 mM Tris-HCl, pH 9.0, 50 mM KCl, 1.5 mM MgCl2, 0.1% Triton X-100, 0.01 gelatine, 200µM of each dNTP, 1 U of Taq Polymerase (Amersham) with 100 ng of each vectors primers. Amplification was initiated by 5 min denaturation at 95°C, followed by 30 cycles of 40 sec denaturation at 92°C and 30 sec annealing

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at 50°C. PCR products were purified through Microcon devices (Amicon, USA) and sequenced using the dideoxy chain termination method on an ABI sequencer (Applied Biosystems, Foster City, USA). The sequences were analysed and alignments performed using the XBAP software of the Staden package, version 93.9 (Staden, 1982). Gaps between sequence contigs were filled by walking with internal primers. *EcoRI* restriction map of cosmids was performed essentially as described in Sambrook et al. (1989).

Northern Blot analysis

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The probes were labelled by random priming with dCTP-(a32P). Hybridisation was performed to human multiple tissue northern blots as recommended by the manufacturer (Clontech, USA).

Analysis of PCR products from LGMD2A families

One hundred ng of human DNA were used per PCR under the buffer and cycle conditions described in Fougerousse (1994) (annealing temperature shown in Table 3). Heteroduplex analysis (Keene et al., 1991) was performed by electrophoresis of ten µl of PCR products on a 1.5 mm-thick Hydrolink MDE gels (Bioprobe) at 500-600 volt for 12-15 h depending of the fragment length. Migration profile was visualised under UV after ethidium bromide staining.

For sequence analysis, the PCR products were subjected to dye-dideoxy sequencing, after purification through microcon devices (Amicon, USA). When necessary, depending on the nature of the mutations (e.g., frameshift mutation or for some heterozygotes), the PCR products were cloned using the TA cloning kit from Invitrogen (UK). One µl of product was ligated to 25 ng of vector at 12°C overnight. After electroporation into XL1-blue bacteria, several independent clones were analysed by PCR and sequenced as described above.

The invention results from the finding that the nCL1 gene when it is mutated is involved in the etiology of LGMD2A. It is exactly the contrary to what is stated in the litterature, e.g. that the disease is accompanied by the presence of a deregulated calpaïn. Identification of nCL1 as the defective gene in LGMD2A represents the first example of muscular dystrophy caused by mutation affecting a gene which is not a structural component of muscle tissue, in contrast with previously identified muscular dystrophies such as Duchenne and Becker (Bonilla et al., 1988), severe childhood autosomal recessive (Matsumara et al.,

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1992), Fukuyama (Matsumara et al., 1993) and merosin-deficient congenital muscular dystrophies (Tomé et al., 1994).

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The understanding of the LGMD2A phenotype needs to take into account the fact that there is no active nCL1 protein in several patients, a loss compatible with the recessive manifestation of this disease. Simple models in which this protease would be involved in the degradation or destabilisation of structural components of the cytoskeleton, extracellular matrix or dystrophin complex can therefore be ruled out. Furthermore, there are no signs of such alterations by immunocytogenetic studies on LGMD2 muscle biopsies (Matsumara et al., 1993; Tomé et al., 1994). Likewise, since LGMD2A myofibers are apparently not different from other dystrophic ones, it seems unlikely that this calpain plays a role in myoblast fusion, as proposed for ubiquitous calpains (Wang et al., 1989).

All the data disclosed in these examples confirm that the nCL1 gene is a major gene involved in the disease when mutated.

The fact that morbidity results from the loss of an enzymatic activity raises hopes for novel pharmaco-therapeutic prospects. The availability of transgenic models will be an invaluable tool for these investigations.

The invention is also relative to the use of a nucleic acid or a sequence of nucleic acid of the invention, or to the use of a protein coded by the nucleic acid for the manufacturing of a drug in the prevention or treatment of LGMD2.

The finding that a defective calpain underlies the pathogenesis of LGMD2A may prove useful for the identification of the other loci involved in the LGMDs. Other forms of LGMD may indeed be caused by mutations in genes whose products are the CANP substrates or in genes involved in the regulation of nCL1 expression. Techniques such as the two-hybrid selection system (Fields et al., 1989) could lend themselves to the isolation of the natural protein substrate(s) of this calpain, and thus potentially help to identify other LGMD loci.

The invention also relates to the use of all or a part of the peptidic sequence of the enzyme, or of the enzyme, product of nCL1 gene, for the screening of the ligands of this enzyme, which might be also involved in the etiology and the morbidity of LGMD2

The ligands which might be involved are for example substrate(s), activators or inhibitors of the enzyme.

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The nucleic acids of the invention might also be used in a screening method for the determination of the components which may act on the regulation of the gene expression.

A process of screening using either the enzyme or a host recombinant cell, containing the nCL1 gene and expressing the enzyme, is also a part of the invention.

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The pharmacological methods, and the use of nucleic acid and peptidic sequences of the invention are very potent applications.

The methods used for such screenings of ligands or regulatory elements are those described for example for the screening of ligands using cloned receptors.

The identification of mutations in the nCL1 gene provides the means for direct prenatal or presymptomatic diagnosis and carrier detection in families in which both mutations have been identified. Gene-based accurate classification of LGMD2A families should prove useful for the differential diagnosis of this disorder.

The invention relates to a method of detection of a predisposition to LGMD2 in a family or a human being, such method comprising the steps of :

- selecting one or more exons or flanking sequences which are sensitive in said family;
- selecting the primers specific for the or these exons or their flanking sequences, a specific example being the PCR primers of Table 3, or an hybrid thereof,
 - amplifying the nucleic acid sequence, the substrate for this amplification being the DNA of the human being to be checked for the predisposition, and
- comparing the amplified sequence to the corresponding sequence derived from Figure 2 or Figure 8.

Table 2 indicates the sequences of the introns-exons junctions, and primers comprising in their structure these junctions are also included in the invention.

All other primers suitable for such RNA or DNA amplification may be used in the method of the invention.

In the same way, any suitable amplification method: PCR (for Polymerase Chain Reaction ®) NASBA ® (for Nucleic acid Sequence Based Amplification), or others might be used.

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The methods usually used in the detection of one site mutations, like ASO (Allele specific PCR), LCR, or ARMS (Amplification Refactory Mutation System) may be implemented with the specific primers of the invention.

The primers, such as described in Tables 1 and 3, or including junctions of Table 2, or more generally including the flanking sequences of one of the 24 exons are also a part of the invention.

The kit for the detection of a predisposition to LGMD2 by nucleic acid amplification is also in the scope of the invention, such a kit comprises a least PCR primers selected from the group of :

a) in those described in table 1

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- b) in those described in table 3
- c) those including the introns-exons junctions of Table 2.
- d) derived from primers defined in a),b) or c).

The nucleic acid sequence of claim 1 to 3 might be inserted in a viral or a retroviral vector, said vector being able to transfect a packaging cell line.

The packaging transfected cell line, might be used as a drug for gene therapy of LGMD2.

The treatment of LGMD2 disease by gene therapy is implemented by a pharmaceutical composition containing a component selected from the group of :

- a) a nucleic acid sequence according to claims 1 to 4,
 - b) a cell line according to claim 24,
 - c) an aminoacid sequence according to claims 5 to 9.

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CLAIMS

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1. A nucleic acid sequence comprising :

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- 1) the sequence represented in Figure 8; or
- 2) the sequence represented in Figure 2; or
- 3) a part of the sequence of Figure 2 with the proviso that it is able to code for a protein having a calcium dependant protease activity involved in a LGMD2 disease; or
 - 4) a sequence derived from a sequence defined in 1), 2) or 3) by substitution, deletion or addition of one or more nucleotides with the proviso that said sequence still codes for said protease.
 - 2. A nucleic acid sequence that is complementary to a nucleic acid sequence according to claim 1.
 - 3. A nucleic acid sequence comprising in its structure a nucleotidic sequence according to claim 1 or 2, under the control of regulatory elements, and involved in the expression of calpaïn activity in a LGMD2 disease.
 - 4. A nucleic acid sequence encoding the aminoacid sequence represented in Figure 2.
 - 5. An amino acid sequence which is coded by a nucleic acid sequence according to claims 1 to 4, characterized in that it is a calcium dependent protease enzyme belonging to the calpain family, involved in the etiology of LGMD2.
 - 6. An aminoacid sequence according to claim 5 or 6, characterized in that either it contains the sequence such as represented in Figure 2, or the amino acid sequence of Figure 2 modified by deletion, insertion and/or replacement of one or more amino acids with the proviso that such aminoacid sequence has the calpain activity involved in LGMD2 disease.
 - 7. An amino acid sequence according to claim 5 or 6, characterized in that LGMD2 is LGMD2A.
- 8. A host cell unable to express a calpaïn enzyme activity, characterized in that it is transformed or transfected with a nucleic acid sequence comprising all or part of the nucleic acid sequence according to any one of claims 1 to 4.

9. Use of a nucleic acid according to one of claims 1 to 4 or a host cell according to claim 8 in the manufacturing of a drug for the prevention or the treatment of an LGMD2 disease.

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- 10. Use of an amino acid sequence according to claims 5 to 6 in the manufacturing of a drug for the prevention or the treatment of an LGMD2 disease.
 - 11. Use according to claims 10 or 11, characterized in that LGMD2 is LGMD2A.
 - 12. Use of an amino acid sequence according to claims 5 to 7 for the screening of the ligands of said amino acid sequence, said ligand being selected in a group consisting of substrate(s), co-factors or regulatory components.
 - 13. Use of a nucleic acid sequence according to one of claims 1 to 4 in a screening method for the determination of the components which may act on the regulation of gene expression of calpaīn.
- 15 14. Use of an host cell according to claim 8 in a screening method for the determination of components active on the expression of the calpain.
 - 15. A method for detecting of a predisposition to a LGMD2 disease in a family or a human being, such method comprising the steps of :
 - selecting one or more exons or their flanking sequences of the gene,
 - selecting primers specific for these exons, or their flanking sequences, or an hybrid thereof,
 - amplifying the nucleic acid sequences with these primers, the substrate for this amplification being the DNA of a human being; and
- comparing the amplified sequence to the corresponding sequence derived from Figure 2 or Figure 8.
 - 16. The method according to claim 15, characterized in that the primers are those selected from the group of :
 - a) those described in Table 1;

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- b) those described in Table 3; and
- c) those including the introns-exons junctions of Table 2;
 - d) those derived from the primers in a), b), or c).
- 17. The method according to claim 15 or 16, characterized in that LGMD2 is LGMD2A.

18. A kit for the detection of a predisposition to LGMD2 by nucleic and amplification characterized in that it comprises primers selected from the group of :

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a) those described in Table 1;

- b) those described in Table 3; and
- c) those including the introns-exons junctions of Table 2;
- d) those derived from the primers in a), b) or c).
- 19. Use of a host cell according to claim 8 in a manufacturing of a drug for gene therapy of an LGMD2 disease.
- 20. Pharmaceutical composition for the treatment of an LGMD2 disease characterized in that in contains a component selected from the group of :
 - a) a nucleic acid sequence according to claims 1 to 4,
 - b) a host cell according to claim 8,
 - c) an aminoacid sequence according to claims 5 to 7.

B) EcoRI restriction map 10-16kb **SUBSTITUTE SHEET (RULE 26)**

F16.1

C) Cosmid map

3A4

163

1911x9Aeq

188

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-641 -601 -481 -361 -241 atatcagitagcciggiticncintacagincatcattitgcitaaagicacagcitacgagaacciatcgatgitaagigaggattitcicigcicag gtgcæc<u>itititititittitte</u>aga<u>sogagicicioicaccigogcicgagigocgigaicigggiicaciacaectcicccicqggiicaagcaalicticigicica</u> OCCICCCAAGLAGCIGGGALIACAGGCACCCCCCCCCCACACCIRALILILIAGIAGAGACAGGGKIRCACLALIGICGCCALGCLGGICLCGAACLCGLGACCLCALG <u>toatccacccoctro</u>gcet cceaagtgcagattagagacgtgatccacatggcccagcaggaccacttttagcagattcagtcccagtgttoattttgtggatgggagagacaa gaggtggcaaggtcaagtggcaggtagagaggattttctcaaatgaggactctgctgagtagcatttccatgcagacatttccaatgagcgctgacccaagaacattctaaaaa gataccoan<u>ictan</u>cattgaataatgttotgatatoctaammattttaggactaamm<u>atcatg</u>ttetetammattcacagamttttgtagamttcagtmootecogttcacctamot Agctitit (graathit gitticcaticatit gaigg ceaglagtigg gict of at anchige teataacaigica geaglic teaglitic titice agig tite accitacica gatactocoliticaliticigocaccaccagcact<u>ical g</u>gcaacagaaalgiccolagocaggileteicicicicalgeagicicicalactoacagigilicitoaca Cagasatcettageacteatteceaggagasettatgetteagaateaengeteggttittaagatggacataacetgteegaeettegatgggettteaaetttgaactggatgt ggacaetttteteteteagatgacagaattaeteeaaetteeeettgeagttgetteettgaaggtagetgtatettattteltaaahagettttetteeaaageeaettge totatttttagttitooiggeicaageaicticaggeeactjaaacacaaceicacicititeteteteeciciggeatgotgetgetgetgagagaceecaagteaacattgett

FIG. 24

FIG. 2B/1

3/33 250 270 270 350 350 350 270 290 290 310 310 330 330 350 350 P D E T S L F X S Q K F P I Q F V W K R P P E I C E N P R F I I D G A N R T D CAMGITCTITATGTGGACCCTGAGTIC

K V L Y V D P E F ATCTGTCAAGGAGGTAAGGAACTGCTGGTGCCTGACCTGAACCAGCACTTGTTTTCCAAGTCATAACCATGATTTCCAAGTTTCATCGAAAGTTTCATCGAAAACTACCAA ATCAGGGATGCTCCTAGTGACATGTACAAGAAAGGCATGGAGGCTCCCTCATGGGCTCCTTCATGATGACAACATGACAACATGGAACATGGAACATTGGAACATGTCCTTCTGGT I R D A P S D H Y K I H K K A I E R G S L H G C S I D D G T N H T Y G T S P S G 10% 10%

SUBSTITUTE SHEET (RULE 26)

۵ L <

H F R A F E

CAGCTCTATGACATCATTACCATGCGGTACGAGACAAACACATGAACATTTGACAGTTTCATCTGCTTCGTTAGGCTGGAGGGCATGTTCAGACCTTTTCATGCATTTGAC

Q L Y D I I T M R Y A D K H M N I D F D S F I C C F V R L E G M F R A F H A F D

ANGATGAAATGATATCAAGCTCAACGTTCTGAATGACTGCAGCTCACCATGTATGCCTGA

2450

2350

FIG. 2B/2

4/33 2050 GTGANCANACACAGGACCTGAAGACACAGGGTTCACACTGGCGTAGCGTAGCATGATTGCGCTCAGATGGCTCTGGAAAGCTCAACCTGCAGGAGTTCCACACCTC V N K H K D L K T H G F T L E S C R S H I A L H D T D G S G K L N L Q E F H H L

SUBSTITUTE SHEET (RULE 26)

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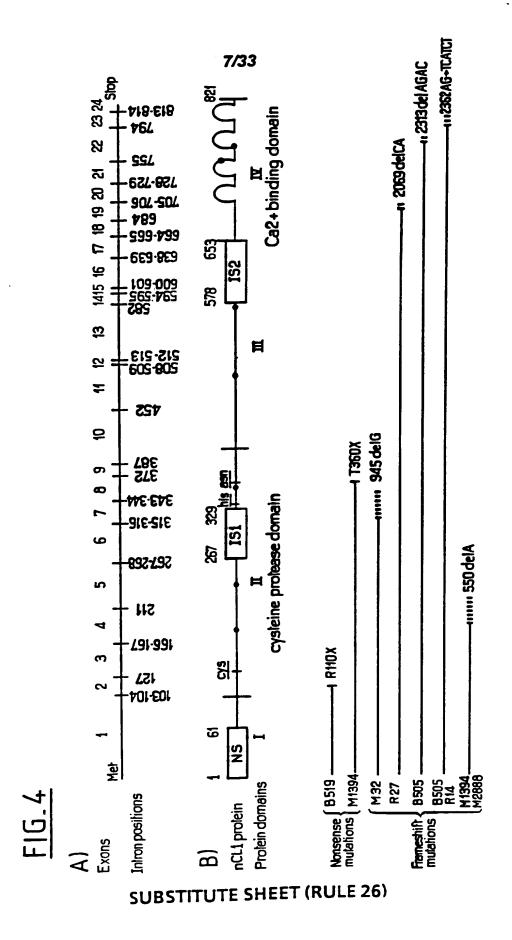
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120 481 481 481 120 120 120 120

FIG. 2C

BICENPERILIDEN GROUP GROUP PROCESSATE AGGOURS GCT N. H. G. C. T. H. G. T. T. EV. DER TERROR GROUP GROU
MPTVISASVAPRTAAEPRSPGPVPHPAC BICENPFEIIDENBIDE GOEFGOOF G. B. C. D. C. C. D. C. C. D. C.

SUBSTITUTE SHEET (RULE 26)



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heart
brain
placenta
lung
liver
skeletal muscle
kidney
pancreas

3.6 kb -

FIG. 5

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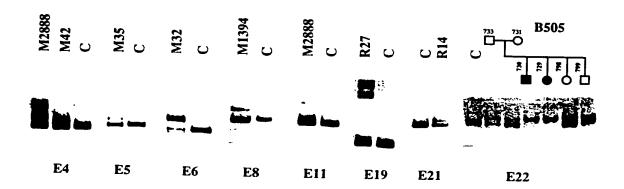


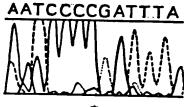
FIG. 6

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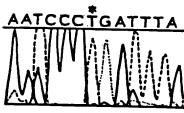
FIG_7

A) EXON 2

Normal sequence

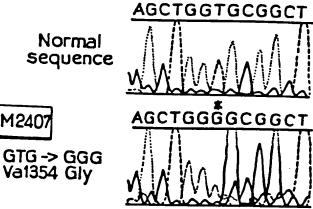


B519



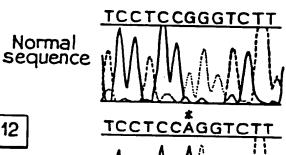
B) EXON 8

Normal sequence



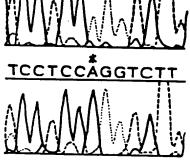
CGA -> TGA Arg 110 Stop

C) EXON 13



CGG -> CAG Arg 572 Gln

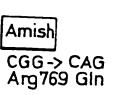
R 12



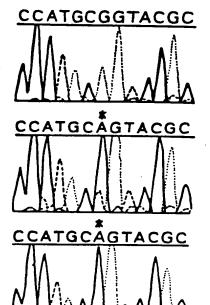
D) EXON 22

M2407

Normal sequence



B 501 CGG -> CAG Arg 769 GIn



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LISTE DE SEQUENCES

(1)	INFORMATION	GENERALE:
-----	-------------	-----------

- (i) DEPOSANT:
 - (A) NOM: AFM
 - (B) RUE: 13, place de Rungis
 - (C) VILLE: PARIS
 - (E) PAYS: FRANCE
 - (F) CODE POSTAL: 75013
 - (G) TELEPHONE: (1) 45 65 13 00
- (ii) TITRE DE L' INVENTION: LGMD GENE
- (iii) NOMBRE DE SEQUENCES: 4
- (iv) FORME LISIBLE PAR ORDINATEUR:
 - (A) TYPE DE SUPPORT: Floppy disk
 - (B) ORDINATEUR: IBM PC compatible
 - (C) SYSTEME D' EXPLOITATION: PC-DOS/MS-DOS
 - (D) LOGICIEL: PatentIn Release #1.0, Version #1.25 (OEB)
- (2) INFORMATION POUR LA SEQ ID NO: 1:
 - (i) CARACTERISTIQUES DE LA SEQUENCE:
 - (A) LONGUEUR: 3018 paires de bases
 - (B) TYPE: acide nucléique
 - (C) NOMBRE DE BRINS: double
 - (D) CONFIGURATION: linéaire
 - (ii) TYPE DE MOLECULE: ADN (génomique)
 - (xi) DESCRIPTION DE LA SEQUENCE: SEQ ID NO: 1:

TGATAGGTGC	TTGTAAACTG	TGCTTAACGA	AAACATACCG	TGTGCTGTAG	GGACTTAACT	60
CTTGTTTATA	TCAGTTAGCC	TGGTTTCGCT	AACAGTACAT	CATTTTGCTT	AAAGTCACAG	120
CTTACGAGAA	CCTATCGATG	ATGTTAAGTG	AGGATTTTCT	CTGCTCAGGT	GCACTTTTT	180
TTTTTTTAA	GACGGAGTCT	CTTTCTGTCA	CCTGGGCTGG	AGTGCAGTGG	CGTGATCTGG	240
GTTCACAACA	ACCTCTGCCT	CCTGGGTTCA	AGCAATTCTT	CTGTCTCAGC	CTCCCAAGTA	300
GCTGGGATTA	CAGGCACCCG	CCGCCACACC	CGGCTTATTT	TTGTATTTTT	AGTAGAGACA	360
GGGTTTCACT	ATTGTTGACC	ATGCTGGTCT	CGAACTCGTG	ACCTCATGTG	ATCCACCCGC	420
CTCGGCCTCC	CAAAGTGCAG	AGATTAGAGA	CGTGAGCCAC	ATGGCCCAGC	AGGACCACTT	480

FIG 8A/1

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TTTAGCAGAT TCAGTCCCAG TGTTCATTTT GTGGATGGGG AGAGACAAGA GGTGCAAGGT 540 CAAGTGTGCA GGTAGAGACA GGGATTTTCT CAAATGAGGA CTCTGCTGAG TAGCATTTTC 600 CATGCAGACA TTTCCAATGA GCGCTGACCC AAGAACATTC TAAAAAGATA CCAAATCTAA 660 CATTGAATAA TGTTCTGATA TCCTAAAATT TTAGGACTAA AAATCATGTT CTCTAAAATT 720 CACAGAATAT TTTTGTAGAA TTCAGTACCT CCCGTTCACC CTAACTAGCT TTTTTGCAAT 780 ATTGTTTTCC ATTCATTTGA TGGGCAGTAG TTGGGTGGTC TGTATAACTG CCTACTCAAT 840 AACATGTCAG CAGTTCTCAG CTTCTTTCCA GTGTTCACCT TACTCAGATA CTCCCTTTTC 900 ATTTTCTGTC AACACCAGCA CTTCATGTCA ACAGAAATGT CCCTAGCCAG GTTCTCTCTC 960 TACCATGCAG TCTCTCTGC TCTCATACTC ACAGTGTTTC TTCACATCTA TTTTTAGTTT 1020 TCCTGGCTCA AGCATCTTCA GGCCACTGAA ACACAACCCT CACTCTCTTT CTCTCCCT 1080 CTGGCATGCA TGCTGCTGGT AGGAGACCCC CAAGTCAACA TTGCTTCAGA AATCCTTTAG 1140 CACTCATTTC TCAGGAGAAC TTATGGCTTC AGAATCACAG CTCGGTTTTT AAGATGGACA 1200 TAACCTGTCC GACCTTCTGA TGGGCTTTCA ACTTTGAACT GGATGTGGAC ACTTTTCTCT 1260 1320 GTATCTTATT TTCTTTAAAA AGCTTTTTCT TCCAAAGCCA CTTGCCATGC CGACCGTCAT 1380 TAGCGCATCT GTGGCTCCAA GGACAGCGGC TGAGCCCCGG TCCCCAGGGC CAGTTCCTCA 1440 CCCGGCCCAG AGCAAGGCCA CTGAGGCTGG GGGTGGAAAC CCAAGTGGCA TCTATTCAGC 1500 CATCATCAGC CGCAATTTTC CTATTATCGG AGTGAAAGAG AAGACATTCG AGCAACTTCA 1560 CAAGAAATGT CTAGAAAAGA AAGTTCTTTA TGTGGACCCT GAGTTCCCAC CGGATGAGAC 1620 CTCTCTCTT TATAGCCAGA AGTTCCCCAT CCAGTTCGTC TGCAAGAGAC TCCGGTGAGT 1680 AGCTTCCTGC TTGCTGGCTG GGTTTCCCCC CCACGGAGGA GTCCTCTCAC TCAGCACCTC 1740 CGGCAGCTCA GCTGTGCACA TGGGCACTGG GGGAAGGATC CTGGCAGCAG CTCTGCTGGG 1800 CTCTGTCTTT AAGTGTGAAG CAGGGAGGAG AGGAACAGGT CTCAGATATT TCACCAAATC 1860 TCAGCAAAAT CCAGAGGGAG AGCGCAGGAG GTGGGGTGAT TCTTATGCTC TGGCTCTTTC 1920 TCTCTGAAAA AAAAAAAAA ATCTTGCTTT TTATAAAAGT GGGTGGAACT CAGTTTAATT 1980 CATCCTGTAA AAATAAATAT TCCTTTCTCA GAACAAATTC CAGACAGCCC AGATGTACCT 2040 GTTCGTTTTA ATATTATTCA TCTTGGTAAG ATTATTTCAG TTTCTCTGGC TAAAATCATG 2100

FIG 8A/2

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					A ACCCTAGAAA	2160
					T AAAGAGAGAG	2220
AGACCTAGC	A ATCGCTTTGC	TCCACCTAC	C TCACCTCATA	A AGTGAGGAG	T CAAGGCACAC	2280
TAGAGTGAAA	A TATATCTAGI	GGGCACATGA	CAGAGCCCG	ATTAAAACT	T TGTTTTAGGA	2340
					CACATTCCCC	2400
					GAGAAATATTT	
					GTGAGTAAGA	2460
					GGCATTGAAC	2520
					GCAAGGAGAG	2580
					GCAAGGAGAG GAGACCCAAT	2640
						2700
					GGCTCATGCC	2760
					AGAGTTAGAG	2820
		GAAAACCTAT				2880
					GGGAAGATCA	2940
		GGCAGCAGTG	AGCCGAGATC	ATGCCACTGC	ACTCCAGGCT	3000
GGGTGACAGG	AGTGAGAC					3018

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(2) INFORMATION POUR LA SEQ ID NO: 2:

- (i) CARACTERISTIQUES DE LA SEQUENCE:
 - (A) LONGUEUR: 11451 paires de bases
 - (B) TYPE: acide nucléique
 - (C) NOMBRE DE BRINS: double
 - (D) CONFIGURATION: linéaire
- (ii) TYPE DE MOLECULE: ADN (génomique)

(xi) DESCRIPTION DE LA SEQUENCE: SEQ ID NO: 2:

GATCCACCCG CCTTGGCCTC CCAAAGTGCT GAGATTACAG GTGTGAGCCA CCACGCCCAG	60
CCGACACTGC CCTAACTCTC AAGTTGCATC CTTACTCGAA TAGTATGACA GTGTGGGAAG	120
CAGCATGGGA CAATGTAAAA AGGAGGCATG TTTCTGGCTT CTGCTACTTA CTAGCTGTGT	180
GTCTTTGCAC GAGTTTCTTA ACCTCTCTGG GCCTCAGTTT CCTTATCTGA AAAATAACAA	240
TGATAGTATT CCCTTCACAG GGCCAAATGG AATACTATCA GGAACACTAC ATAATGGAAC	300
TCAATAAATA ATAGCTACTG CGGCCGGGCG CGGTGGCTCA CATCTGTAAT CCCAGCACTT	360
TGGGAGGCCG AGGCGGGTGG ATCACAAGGT CAAGAGATGG AGACCATCCT GGCCAACATG	420
GTGAAACCGT ATCTCTACTA AAGATACAAA AATTAGCTGG GCATGGTGGC GCATGCCTAT	480
AGTCCCAGCT ACTCGAGAGG CTGAGGCAGG AGAATCACTT GAACCCCGGA GGCAGAGGTT	540
TCAGTGAGCC AAGATTGCAC CAGTGCACTG CAGCCTGGCG ACAGAGTGAG ACTCCGTCTC	600
AAAAAAATAC CTATCTATCT ATCTGTCTAT CTACTGTTAT TCTTACCTGG TCATTTCCTT	660
TTTGTTTCAC AGGAAATTTG CGAGAATCCC CGATTTATCA TTGATGGAGC CAACAGAACT	720
GACATCTGTC AAGGAGAGCT AGGTAGGAAA GTGCCTCAGG TCAGATCCTG CCAGATGATC	720
AAGGGGTGAT TACAAGGTGT GATCCCCTTC CAGGAGGTAA AGGGACAATC TGTGCTTGCT	
TCCAGTAACT TTTTGGAAGA TTTTTTATAA CAGTTGCTTT ATGGTCGTTT ATCTACATGC	840
TGGCGATTGC TTCATTTCCT CCTACATGCC TCTTTAGCAC TCTGCCATGC ATCACAGGGG	900
GTATCTGCAT CCTGTGGCCT CCTCTCCAGT ATCTCAAGGA CACTTACATA CCCCACTCAG	960
CATGACAAAA GCCCTGCTTT TCACTGTATC GTCTTTCTTG GAAGACAGCT CTGTGACTGT	1020
GCACCAAGCA TGCCCCTTGG GCATGGAGAT TCTAGATACA CACACAAAAG GCATCGCCAA	1080
GGAAAGCACT TGTAACTGGA ACCCTTGGTT TAAATTGGCC CAGCATAGCT CCATCTTTAA	1140
THE THOUSE CAGCALAGET CCATCTTTAA	1200

FIG.8/B!

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AAGAGTCTTT CCACAAAGAT GGCATCCGCC ATGTGGATGA GCATCCAATT TTCTCTTTGA	1260
TTGGTTAGCT TGACTGCTCC ATCTGATCTT CCTCTCTCTC GACCTCTTGT TCAGAAAGTA	1320
TTGTCTTTGG TGTGGACTAT AAGCAAGCTC TGTGAAGTAA AATTGGAGAG AACACCAACA	1380
GAAACAATTT AAATTTGAGG AAAAGGGGGC ACCTAAGACC AAAGGAATTT GGCTTATTTC	1440
ATTCCAGAAG GGGAGGCTGA GAATAAATCA GATGAATATC TGGGTTCCTG CACCTGAGGG	1500
AAGGCTTCCT GCAGAGCCCT GGGCATAATA ATCTGGGACC TTCAAACCAA TAACCTCTTT	1560
TCCAAGGAAA GACTGGCTGC TTCCAAGGAG GGTAGGGGAG AGTCGGGCTG CAGGCAGCTC	1620
TCAAGTCTCC CCTTGCACAC TCTCAGGTTG GCATTTTCAC TTTAACCCAT CCTCCCTTAA	1680
GAAGGCAGTT CTTTGTGACC AGGGTACACC CCCTATTATA TATATATATA CACACAGA	1740
GAGAGAGAGA GAGAGAGA GAGAGAAAGA GAGCAAAGTG TTACCTCCAA CTACATACAG	1800
TACTCTGTCA GAAAAGAGGT TCAGAGAATA AGAAAACGTC CCGAGCTCAT TCCGTTGCCA	1860
GCAATGTCTT ACTGCCCCCT ATAGACGGGT TCCAGGGCAG CTGCCTACCT GGCCTTCCTT	1920
CCAATACAAA TCATCTTGGT GGATGGTTCT CTGAGGCTCA GTCTTCGCTG AAGTCAGAAG	1980
AGGAATTGGA CTCACATTGC AAAGGCACAG GGCAGGGCAG	2040
AGAACAACCC AGTTATGATC ACCTACTGCT CTGTCTCCAT TGAGGCCTAA AAAGGAAGTG	2100
AGTTTATACT GCAGTTGGAG GAACTGCCTG CAGCCTTGAG GAAAATGTCT AGTCACAAGG	2160
GAGTAAGTTA CCTGTTGATC ATATTGTCAA GGAATTCCTG TCCAATTCTC CTTCCCTGGG	2220
TTGACACCTC TGTAAGGTCA GATCTGGAAG TAGGAGAGTG GGCACCAAGG GAGTCCCCGT	2280
TCAGGGAAGT GGAGTGGCTG GCTGGGATTG GGGCTTTTTC TTCCCAGGAG GAGCAGGAGT	2340
GCTCACGATC TGTGCCCTGT GTCTGCCTGC AGGGGACTGC TGGTTTCTCG CAGCCATTGC	2400
CTGCCTGACC CTGAACCAGC ACCTTCTTTT CCGAGTCATA CCCCATGATC AAAGTTTCAT	2460
CGAAAACTAC GCAGGGATCT TCCACTTCCA GGTGAGGTAA TGAGAGTGTA GTTAAGAGGG	2520
CCAGCGGCAG GCCACCCACC GCTGGTCTCC TGGCCTTGAC TTCCCAGAAG CTGGAGGAAA	2580
CTTCCCACCC ATCTACCCGC AGCGGCAACA GTCGGCATGG ACCCCCTTAA GGCTTCAAGC	2640
CTGGGAGGAA GCAGTTGCTT ATCTCTGGCT CCCTAATCCC TCCCCCACCA CCTTCCACTA	2700
	2760
GTCTGGCTGC TTTTATTGCC TGCAGCCCTT CTCAAGTAGG TCCCTAAGAT ATTAGCACTG	2820

FIG. 8B/2

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TGACACCACA GGACCCTTCA GGTTGTACAG GAACCCCTGT CCAGGGCTCC TGTATACTTC	2880
TTCCTCTCTA AGGCATGGCG GTACCAAGGC TATCACTCCT CTCTTCCAAG CCCTGGAAGA	2940
AGAGTCTGCT TAACCTGGGG ATCAGGCTTC TTGTTTGCCC TAGAACTGAA TCTGATGGTT	3000
CTAGAATCCA TCCAGCTACT GGAAATTTTC TGGGTCCCAG TCACCTTGGC ATAGAGCTGG	3060
TGCTAGAGCA GAACCAAACT GAATTCTACC TGTGAGGGTC TCGTAGCTTC CGGGATGCTG	3120
GGGAGTCAGC CTGTCTCCAG CTTCAAAGGC TCCCTCATGT CCCAGGATGA CCCACATTAT	3180
CAGTTCTTGC TCCCCGGGTC TTGCACCTCA GCACGGAAGG CCTCAGAAAA GGTCTGTCTC	3240
CAGGCTCAGA CTCCCCCTCC TGCCGCCTTG GGAACATGGC ATATTTAAAG GGTCTCAGAT	3300
CTAAAGGGCC TTACATACAA ATATCAGATA GATTTCTGTT CTCATTTCAA TGAGGGAGAA	3360
AGTGCCATTG AAAAGGAGAC TAAACCACAT TTGGCCCTTT TCAGTTCAAA CTGATTCATT	3420
CAAAAAAGAG CGACATCCAA ACTTGAAATG ATTGAACAAT GTTCCTGCTA CAGCTAGAAT	3480
AGATTCTGGG TCACTTTGTT CCTCCGTTTC AATCCTTGTT CTTCAGTTTG GCATCAAGAA	3540
ATACCTAAAT CAGCACAGTG CCTTCACTGC ATAGTTCCCA ATCCTGGCCA CATTGAATCA	3600
GCTGGGGGCA CCTGAGAGTG CTGACACCCA GGCCCTGCCC CAGACCTGCT GAGCAGGAGA	3660
ATGAAAATCT TACATCCTAA GACACTCATG GAGCACCTAC TCTACCCATT ACTGGGCTGG	3720
ACTCTGTGGA AGACATGAAG TATATGTAAC TCACTTCCAG CTCTCAAAAA GCACCCAGTC	3780
CAGTTAGAGA CAGATTTACA CACCCCAAAC ACAAAATAGG ATGAACAGGC ACCCAGATGC	3840
AGAGTCCAGG AAATGATGCT GCTTTGGGAT TCAAGAACCC CCTGAGGAAT GTGGAGGAAG	3900
GACACATTTC CTAACAGTAA TTTGAGTATG TGACTCTGTG CGTGACGCTT CTGTGCAGTT	3960
CTGGCGCTAT GGAGAGTGGG TGGACGTGGT TATAGATGAC TGCCTGCCAA CGTACAACAA	4020
TCAACTGGTT TTCACCAAGT CCAACCACCG CAATGAGTTC TGGAGTGCTC TGCTGGAGAA	4080
GGCTTATGCT AAGTAAGCAA CACTTTAGAA TGTGAGGTGG GGCTAGAGGT GAGAAAGTGG	4140
GTTGCAAAAT CCAGCCGAGA CCTCACTCAC AGGAAGAGGC ATGTGCCTCT ATACGTGCAT	4200
ATGTGTGGGC ATGCAAGTCC AACTGTGACC CAAAGTTAGA GATCAGTTCC AGGCAACAAC	4260
AGCTCTAACT AAAAACATTA AATTTAAGAG TAGAAATGAA GATTTGCATA GAAGACCTTT	4320
AGCTTTAGCT CACCATAGCG AGTTCTTTCA TTGCACCTCC ATGGTGGCAT TGCAAGTCTT	4380
GGGATCAGAG CATTGTCCCA GGGTCTCGAT TGGCTCAACC TCATGTGCTT ATAGAAGATT	4440

FIG.8B/3

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TATAAAGACA TGTTGTCTC	T CAACTTAAA	A GCTCCACCC	C AGATGATAA	T AATGGATTTT	4500
CAAATTTTGG AACAAGGTC	A CTCTGTAAT	G CAGGCTGGA	G TGCAGTGGT	G CAGTCACGGA	4560
TCACTGTAGA TTGACCTCC	T GGGTTCAAG	G TGCTCCTCC	C ACCTCAGCC	T CCCAAGTAGC	4620
TGGGACTACA TGCGGGCAT	C ACCATGGCC	C TTTTATTTT	CTATTTTTT	T GTAGAGCGGG	4680
GTTTTCCCAT GTTGACCCA	G ACTGTTCTC	G AACTCTTGG	G CTCATACAA	T CCACCAGCCT	4740
TGCCCTCCCG AAGCGCTGG	G ATTGCCGGT	G TGAGCCACCA	A CACCGGCAG	C TGCTAATGGC	4800
TTTAATGCAG CCCTTCCTC	A ACGTTCAGG	A TGTAGTGGAA	AGAGCTCTC	A GGAAGTGGGG	4860
ATAGCTGGGT TTCAATCCC	A GTGCTTCTGC	CTCTCTGTGC	TCTTGGGTG	G GTCACTTAGC	4920
CTCTTGAGCT CAGTTTCTT	CATTATGAAGA	A AAGGGAATCA	TTGTTTCCAT	CCCATGAGCT	4980
CATAGGGTTA ATGTGGAATT	GATGAAAGAA	CATCACAGCA	TCCAAGAGGT	AAAGTTCTGG	5040
TGGCAGTGGT ACCTGGGTTT	TGTTCCCTGG	AACTCTGTGA	CCCCAAATTG	GTCTTCATCC	5100
TCTCTCTAAG GCTCCATGGT	TCCTACGAAG	CTCTGAAAGG	TGGGAACACC	ACAGAGGCCA	5160
TGGAGGACTT CACAGGAGGC	GTGGCAGAGT	TTTTTGAGAT	CAGGGATGCT	CCTAGTGACA	5220
TGTACAAGAT CATGAAGAAA	GCCATCGAGA	GAGGCTCCCT	CATGGGCTGC	TCCATTGATG	5280
TAAGTCTGGG GTGTGGGGCA	CAGGGTGGGG	AGCTCCAAGT	GTCAGGAAGC	CTTTTACCCA	5340
ATGAAGGGCA GCATAGAGCT	TTTGTGTGGG	ACAGAGCGAA	TGTTTTGTTT	GAGGAAGCAG	5400
GAACTGGCTC TCAACTTTGA					5460
TTCAATAAAG ACACTGGTCA					5520
AGAGGCCTGT GTCAGTGGAG					5580
TTAAGCTGGC CACGTACTTG					5640
CAGTCACTGG GCTTTCACCA					5700
GGAGAAGCTA AGCCTGCAGC					5760
CCAAGGGTAG GGAGATGGCC					5820
AGGGCTGCTC AGTATATTGA					5880
GAAGCTGAAT TCCTGCCCCT					5940
TTAGCACACA ACACCATGGA					6000
ATTCGTGCTC TGTTGATCTC	TCCTCTCTCC	CTTTGTCTGT	CCCATCTCTT	тстсстстст	6060

FIG. 8B/4 · SUBSTITUTE SHEET (RULE 26)

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CCTTCCCTTT CCACCCTTCT GTGTTTGTTC TCTCCCTCCC CTGTGTTGTT CCCTACATTC	6120
TCCATCGGGC CTCAGGATGG CACGAACATG ACCTATGGAA CCTCTCCTTC TGGTCTGAAC	6180
ATGGGGGAGT TGATTGCACG GATGGTAAGG AATATGGATA ACTCACTGCT CCAGGACTCA	6240
GACCTCGACC CCAGAGGCTC AGATGAAAGA CCGACCCGGG TGTGTACACC TCCGATTATC	6300
AGAACTGACC ATCCCTCCAA CCCACATGAC CCCGCCCTAT TAGTGTCAGA CTCCCCTCAG	6360
CAGCCAGGGC CTTACCCACA CACCCCCACC TGGCACCTCC CAAGGGTCTG GGTTGAAATA	6420
ACTTGCTCAG CCAAGGCTCC TGAAGAGGGT GCAAGAACCA GGATTTTGGA GGGAATCTCT	6480
GCTGGAGTTT CTGCATATTC CATGGTCCAG GCAGTTCCTC TCATAACGAA CTATGAGACA	
GAAATACTTG TAAAGATACT TCATTTATTT TGAAATATTT TTCCTCTTCT AATGTATTCA	6540
TTTATTCATT CAACACTTAT TTTTGAGCTC CTACTATGTT CCAGGCACTC CTCTAGCAAA	6600
CAAAGCAAAT TCTCTCCTCT TTTTCAATAT TTGTGGAAAA AGCAAGGTCT CCCTCTTGTA	6660
GAGTTTATAT TCTAGTATTT TCATAAGTTA TACCTGCTCA CTGGAGAATA CTGAGCCATA	6720
CAGAAAAACA CAGAGGAAAA TTTCACTTAT ATTTTTCCCC ATGTAAAGAT AACCACTCTT	6780
AACATCTAGT ATATGTTCTT CCAGGATTTT TCTATGCACA CACTGAATCT GTATTTTAT	6840
TTTTAAAATG TTATCATATT GTATGTACCT CTTTGCAGCC TGCTTTTTTC AGTTAGTTTT	6900
TTTGGTTTTT TGGTTTTTTT TTTTTTTTGG AAACCAAGTC TTGCTCTATT CCCTAGGCTG	6960
GAGCACAGTT GTTGCCATCT CGGCTCACTG CAACCTCTGC CTCCAAAGTT AAACTAATTC	7020
TCCTGCCTCA GCCTCCCGAC ATAGCTGGGA TTACAGGCAC ACACCACCAC ACATGGCTAA	7080
TTTTTGTATT TTTTAGTAGA GACGGGGTTT CACCATGTTG GCTGGAATGG TCTTGAACTC	7140
CTGACCTCAA GTGATCCACC TGCCTCAGCC TCCCAAAGTG CTGGGATTAC AAGTGTAAGC	7200
CACCACACCC GGCCTAGTTT GATATTCTTA ATGTGCCCAA AGTATTCTCC TGTAACATTT	7260
TTTAATAGCT ACACAATATT CAAACACACA GATATGTTAT AATTTATTTA CCCAATACCC	7320
	7380
TATTATTGGA AAGTTGAGTT CTTTTTTTC TTTGTTTTGT	7440
AAATGCTATA ACGAACATCC CAATAGATAC ATCTTTGTAT ACATCCATGG TGACTTCCAT AGGACAGATT CCCAGCAGTA CAATTCCTCC CTTCAATTCCTCC	7500
AGGACAGATT CCCAGCAGTA GAATTGCTGG GTTGAATGAT ATGCTTAGGG TAATGACAGA	7560
AGAGTCATTT CAAGCAGCTT CCTAGGGTCT TAGAACTTAA GGATTAATGA GTCTTCCCGC	7620
CCCCTCCCAG TCTATTCAGC ATGATCTGGA TCATGAGGAC TGAGATCTGG AAGAGACTGA	7680

FIG. 8B/5

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CATCTCCCAC ACCOTO : 5	
GATCTGGGAG AGGCTGAGAT ACCAAAAGCC CTGGCTCCAC CCATACCCCT CGCCCTGAAA	7740
ACAGCTCTAG GAATTCCGCG GCCTAGCAAG GCTCCGGGAA GCTCCTTTTA AAGCTGTGAC	7800
GTTAGTAGGC ACATGGACCA TAGAGACCTA TCCAGGGCTC ATGGGACTTT AGTGATCCTG	7860
CCCTTCTCCC AAGGATCCCC CATGGCTGCA ACTTGGAAAT TTCTGCAAAT GGAAGAGCTA	7920
CTCCTTAGGC ACGGTCATGT CTGAGCAGGG ATCTCCTCGG GCTTTCTTAG AATTCTCTCC	7980
CTGGGCACTG GGACTCTTGA TTTCTTGAAT ATTATGTTCC AGGTGGGTGT GGAGGAGGTG	8040
AGGGGATGTA AAGAAGGCTA GACTTGGCCA GGCGCAGTGG CTCATGCCTG TAATCCCAGC	8100
ACTTTGGGAG GCTGAGGCGG GTGGATCACC TGAGGTCAGG AGTTCGAGAC CAGCCTGGCT	8160
AACATGGTGA AACCCCGTTT CTACTAAAAA TACAAAAAAT TAGCTGAGCA TGGTGGCACG	8220
TGCCTGTAAT CCCAGCTACT CGGGAGGCTG AGGCAGGAGT ATCGCTGGAA CACGGGAGGC	8280
AGAGATTGCA GTGACCCGAG ATCGCGCCAC TGCACTCCAG CCTGGGCGAC ACAGCAAGAC	8340
TCTGTCTCAA AAAACAAAAA AGAAAGAAAA AAAGGAAAAG CTAAGACTTA CATGTGTCAC	8400
TTAACCCCTT TTCTCAAACC TCTTTCTCTT CCAGGAATAG TCAACCCCTG GATGGCTTCA	8460
GGGGAAGGGG GATCCTGAAG CCCAGGGCAG CCTCCAACTC TACCCCTTCC TCCTTTGAAG	8520
GATACTAAGG GGTCCAGAAA GGAGGGGCAG GACACTGTTA CCCACCCCAC	8580
CCACATTGCT CTCTGATGCT CAGGACAGAG CCTTCTCAGG GAGACCAGCC TGTCTGGAGC	8640
TGTGTCTCTT GGCACTCTTA AAGGGCCACT GAAGGTCCGT TCGTGGTCGT GAGGCACACT	8700
TTCAGGGAGC AGAGTGGTCT GTGTCTTCAC AGAGCCCGGA AAATGAACTA GTATGAACTT	8760
TGCCTCCAAG CAGCAGAACT TCTGTTCCCC CGCCCCTAAT GGGTTCTCTG GTTACTGCTC	
TACAGACAAT CATTCCGGTT CAGTATGAGA CAAGAATGGC CTGCGGGCTG GTCAGAGGTC	8820
ACGCCTACTC TGTCACGGGG CTGGATGAGG TAAGCCTGGT GGGGCTTGGT GGGGCAAGGG	8880
CACCCTCCTG GGTTAACCTC ATGAAGTCAG GACTTAGCTG TTGGGGCCCC TGCCCTGTCT	8940
GCAGAGCTTG CCTCCAATCA GGACATTCAG TTCAAGGTCC AAGCCACGCC TGGGAGCAGA	9000
GGGGCCTGTG AAACTGGTAG AGGTGGATCC TGCCACAGTT GGTGCACAGT TTATCTTTGC	9060
TTTTCGTGCT AAAGATGGCA ATTTTTCCAA CATTTCCAAT GAACAAATTG AAATATCACT	9120
TAACTTTGCT TTTACAAAGT TGGTTTCATG TGTTCTTGAG CTTCCTGTTC TCTCGTGTTC	9180
	9240
AGATAGCTAC AGTTGTCTCT GGGTAGCCAC GGGGACTGGT TCCAGAAGCC CCAACAGTAA	9300

FIG.8B/6

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CAAAATCTGC AGATGCTCAA GTCCCTTCTG TAAAATGGAG TAGTATTTGC ATATAACCTA	9360
TGCACATCCT CCCATATACT TTAAGTCATC TCTGGATTAC TTACGATACC TAACACAATG	9420
GAAATGCTAT GTAAATAGTT ATTGCACTGC ATTGGGTTTT TTTGGTATTA TTTTCTGTTG	9480
TTGTATTATT ATTTTTCTT TTTTTGAATA TTTTTGATCC ACAATTGGTT ATATGCCAAA	9540
GCCATGGATA CGAGAGGCTG ACTGTTCTGT TTTGCTCCTT CTGGGACTTC TGGGTTTTCC	9600
TGGACCATGT CTGAGACAGG AACGTTGTAA GACCTGTTGC ACACAGTTGG GCAGGTTGTG	9660
CCCTGTACAG AGGGATGGGC TGAGAGGGGC AGTTGCCTGC ATCACCCATT GCAGCAGACT	9720
GGAGGGAGTC TGCTTGTTTG TAGTTCCTCA GTCAGCAGGG GCCTTTTGTC TTTCCTTCCT	9780
TTCCTTTTTT TTTTTTTTG AGACGGAGTC TCACTCTGTT GCCCAGGCTG GAGTGTAGTG	9840
GCACAGTCTC GGCTCACTGC AATGTCCGCC TCCTGGATTC AAGCGATTTT CCTGCCTCAG	9900
CCTCCTGAGT AGCTGGGATT ACAGGCGCGT GTCACCATGC CCAGCTAATT TTTGTATTTT	9960
TAGTAGAGAT GGGGGTTTCT CCATGTTGAT CAGGCTGGTC TCGAACTCCT GACCTCGTGA	10020
TCCGCCCACC TCGGCCTCTC AAAGTGCTGG GATTACAGGC GTGAGCCACC ACGCCTGGCC	10080
AGCAGGGGCC TTTTTTCTAA TTTATATGAA GACACCTAAT TTATATGTGT TAGCAAAGCC	10140
CTCCTGTTTA TGCCTCACCT CCTCCCCGA AGCTCATACG GCAGGATGTT CCTGAGAAAA	10200
TTGCCTCTTA GAAGATAGAG AGGAGATGCC AAGCCTAAGT TAGGCAGACT CAGGAGGATA	10260
GGTCTGACCC ACCCCCTGCC ATTCCCCAGC ACACTTGTGA TTAATCTCCT TGGCCAGAGC	10320
CAGGCAGAAC ACCCTCGCGT AAGAGATTTG CCCCCCAGCC CCGTCCCAGC CCTCAGCTAG	10380
ACAGAAGATT CCCTTTCCAG AGAGGCTGCA GAGCATGAGA GCTCTTTCTG TGTGCTTAAG	10440
GTCCCGTTCA AAGGTGAGAA AGTGAAGCTG GTGCGGCTGC GGAATCCGTG GGGCCAGGTG	10500
GAGTGGAACG GTTCTTGGAG TGATAGGTAG GTGAGGGGAC CCCACGGGAT TGGCGGTGGC	10560
GGGGAACAGG GTCCGGGACA AGGCTGTGTT GGGAACTGAG CCATGAGAGT ATTGAAGATG	10620
CTTGGTATAA AATCACCCTC AAAACCAATG ATCCGCAGAG AAGAGGGGCA CAGGTGTTGG	10680
CTCCAGGGAA GGGCCAGGAG TGGAAGCGGG GTGCTGGGGA CCCAGAGAGG TTGCTGACAA	10740
CCATTGGCTG GAAAGGAAGG ATTCCAGAAA GCGTGGGGAA GGTCCAGGCA GGAAAAGCGT	10800
	10860
TCCTGTGAAA TGGGAACAGT ATTATTAGCA CTTACCTTGT GGGCTGATAT TGAGGAGTAA	10920

FIG.8B/7

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CTGGGACTT	TTTTTCCC					
	o iiiiiddd	AGTGCTGAG	CATTGCTAAG	ATTCCCCTTA	CCCGTGCTTG	10980
TCCCTTGTAT	TAAGGCACAA	GGGCCCTTTG	AAAAGAATTT	TACCTGCTTT	ATCAATTGAA	110.0
AGGGATTAAG	ACCTTGGGG	CCAACCCAAA	ATA		TTTATAGGCT	11040
CCATCCACAC	TTOOTH	- 0.2.000/244	HIMAMCAIGC	GAACTTATTA	TTTATAGGCT	11100
CONTROLACAC	, ITCGTAAAAC	CTCCATGGTC	CTACTGGTTC	CTGATTACCT	CCACTCAATG	11160
AGAGGCAATT	CATTACTGAA	TGAGCCATAA	GCGCCTCTTA	TTTCGACACC	GGGATGGCAG	
GACTCAGTCG	AGGAGAAGGA	CCCC+CCC+C		TTTOORGAGG	GGGATGGCAG	11220
TTT 4 0 = 0 = 0	CIICIEIGOA	CCGCACCCAG	GCAGCCTGGG	CCCCTCGGCT	CCTGTACTTA	11280
TTTACTGCTG	GGTACTTCCT	AGCCCAGCAT	GTAATTACTG	GTTCGTTCAG	TCATTCCTTT	112/0
AGTAAATGTT	TCTTGGGCAC	CTACTACATA	GGACCCACAG	Ome.	-0.11100111	11340
CTTTCTACCC	A CCCCC		GONGGCACAG	GTCAAGGCAC	TGGGGATATT	11400
or restrict	ACCCCCTCCC	TCCCTACACT	GTGATTAGGG	ACTGACCGAT	С	11451

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(2) INFORMATION POUR LA SEQ .ID NO: 3:

- (i) CARACTERISTIQUES DE LA SEQUENCE:
 - (A) LONGUEUR: 1834 paires de bases
 - (B) TYPE: acide nucléique
 - (C) NOMBRE DE BRINS: double
 - (D) CONFIGURATION: linéaire
- (ii) TYPE DE MOLECULE: ADN (génomique)

(xi) DESCRIPTION DE LA SEQUENCE: SEQ ID NO: 3:

ATTITITIT TITTITIGA GACGGAGTCT CACTCTGCCA CCCAGGCTGG AGTGCAATGG	60
CGCGATCTTG GCTCACTGCA ACCTCCGCCT CCCGGGTTCA AGTGATTCTT CTGCCTTAGC	120
CTCCTGAGTA GCTGAGACTA TAGGTGCCCG CCACCACGCC CAGCTAATTT TTGTATTTTT	180
ATTAGGACGG GGTTTCACCA TATTGGCCAG GCTGGTCTCG AAATCCTGAC CTTGTGATCC	240
GCCCACCTCG GCCTCCCAAA GTGCTGGGAT TACAGGTGTG AGCCATTGCG AGCAGCCCAG	300
AACTCAATTC TTAACCTTTA AAGTATGATG AGAAGAAGGA TCAAGCCCTC ACCAGCCCAT	360
TTAAGGAGTT TAGGCTCAGT CTTGAGGATG TGAGAAGTCA TTGCTATTGG GTTTCACACT	420
GAGGTTAACA GGTGAAGTCA GCATTTTGGT AGTTCACAGC AGCTGCAACT CTTTGTATTT	480
CTCTGATACC TCCTGTCCCA ACCTACATCA GGCCTTCCCT TCTTCCTGCT TCCTTAATTC	540
CTCCATTTTC CCACCAGATG GAAGGACTGG AGCTTTGTGG ACAAAGATGA GAAGGCCCGT	600
CTGCAGCACC AGGTCACTGA GGATGGAGAG TTCTGGTGAG TCCAGAACCC AGGAAGACCC	660
AGAAGGGTAA GGGTGGGGAA GAGAGGGGAA ATCTCAGACC TCAGTCCCCA GCTAAGGTTA	720
TCAGATTCCA GCCCTTGGGA GATCTTGGCT GTGTTCTCCT CCAGCCCAAG GCCCAGCAAG	780
GATGAGGTTC TGAGAGGAGC CTTCCAGGCC ACAGGGACAA TGAGCCCAGG ACCAGGCCAA	840
CATGACATGG CTCTTGCCTC CTGTGTGCCC CTCCGCCACA CACTCTATTC CAGCCACAGG	900
CACCCTGGCC TTAGCACAAT TCTTTTCTGA GCCTAGGAAG CTCCACTTAC CCTGATCTTC	960
CAACGTCAAC CTCACCCTCT CTCAGGTTGT TTCTATTCAG GCTTCAAGTC TCAGCTTAAG	1020
GAGAATTTTC AAGTCTCAGC TTAAGGAGAG CCCCCTAAGT TCCCCGAGGA CTGGGATTAA	1080
TTTATGATGC TCATCACCCT TAAAATTGTT TGCTTAAGCC GGGCGCGGTG GCTCACGCCT	1140
GTAATCCCAG CACTTTGGGA GGCCGAGGTG AACGGATCAC GAGGTCAGGA GATCGAGAAC	1200

FIG. 8C/1

WO 96/16175	
WO 30/101/5	PCT/EP95/04575

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ATCTTGGCTA	ACACGGTGAA	A C C C T C T C T C T				
	- HOHOGGIGAA	ACCUTGTCT	G TACTAAAAA	ACACAAAAA	A AGTAGCCGGG	1260
CGTGGCAGCG	TGCGCCTGTA	GTCCTAGCTC	CTGGGGAGG	TGAGGCAGGA	A GAATCACTTG	1320
AACCTGGGAG	GCAGAGGTTA	CAGTGAGCCC	AGATTGCGCC	· ACTCCA CTCC	AGCCTGGGCG	1320
ACAAGAGAGA	CTCTCTCT		1101111100000	, MCIGCACICC	AGCCTGGGCG	1380
	CICIGICITG	GAAAAAAAA	AAAAAATGTG	GTCTTAGTTT	AATGTCAAGG	1440
GAAAGGTTTT	GGGTGTTTTT	ATTACTTTAT	TTTTTATTTA	AAAACTATAA	TAGAGACGGG	
CCTCGCTATA	TTTCTCGGGC	TCCTCTCAAA	OD o o o o o o o o		INONGACGGG	1500
000000		TOGICICAAA	CTCCTGGGCT	CAAGCGGTCC	TCCCACCTTG	1560
GCCTCCCAAA	ATGCTGGCAT	GTGGGCCTGG	TCAACATATG	GGACCCCAAC	TCTACAAAA	1620
TAAAATTTA	TAGCCAGATG	TGGTGGCGTG	TCCCTCT+CT		TGGGAGGCTG	1020
A A C C A C C C C C C		1001000016	IGCCIGIAGT	CCCAGCTACT	TGGGAGGCTG	1680
MAGCAGGGGĢ	TCACTTGAGC	CCAGGAGGTT	GAGGCTGCAG	TGAACTATGA	TTGTCGTTCA	1740
CTTTTCTTCT	GAACGTGAGA	TTAAGTGTAG	TCAGCAATTT	CCCTTA CO	TATTTATTCA	2740
GAATTTTTA A	CCCTCACCC		OAATTI	GGCTTAGGAT	TATTTATTCA	1800
	CCGTCACGTT	GCGGCAAACC	AGGT			1834

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(2) INFORMATION POUR LA SEQ ID NO: 4:

- (i) CARACTERISTIQUES DE LA SEQUENCE:
 - (A) LONGUEUR: 14664 paires de bases
 - (B) TYPE: acide nucléique
 - (C) NOMBRE DE BRINS: double
 - (D) CONFIGURATION: linéaire
- (ii) TYPE DE MOLECULE: ADN (génomique)

(xi) DESCRIPTION DE LA SEQUENCE: SEQ ID NO: 4:

	AGGAGGTGGA	GGTTGCAGTG	AGCCAAGAT	CATGCCACTG	C ACTCTAGCC	T GGGCAACAGA	60
	GCGAGACTCT	GTCTCAAAAA	ATACACACA	CACACACACA	CACACACACA	C ACACACACAC	120
	ACACACATAT	ATATACACAC	ATATATATA	ACACACATAT	CACACACACA	CACGTCTGTAT	180
	ATATATGTGT	GTGTGTATAT	ATACACACA	ACACTATTCT	ATATATTCTT	GTAGAGCTAT	240
	GTGTGTCTCC	TGTGCTATTG	AGCATGAGCC	CTTTTTTTT	TITITITI	TTGAGACAGA	300
	GTCTCACTTT	GTCGCCCAGG	CTGGCATACA	ATGGCGCAAT	ATCGGCTCAC	TGCAACCTCC	360
	GCCTCCTGGG	TTCAAGTGAT	тстсстссст	CAGCCTCCCA	AGTAACTAGG	ATTACAAGTG	420
	CCCGCCATAA	TGCTCAGCTA	ATTTTTGTAT	TTTCAGTAGA	GATGGGGTTT	CACCATGTTG	480
	GCCAAGCTGG	TCTCAAACTC	CTAGCCTCAG	GTGATCCACC	TGCCTCAGCC	TCCCAAAGTG	540
	CTGGGATTAC	AGGCATGAGC	CACAGCACCC	TGGTGAGCAC	TAGAGCTTAT	TTCTTCTATC	600
	TAACTGTATT	TTTGTATCCA	TTAGCCACCC	TCTTTTCATC	СТССССТСТС	CTTCCCTTCC	660
	CAGCCTCTGG	TAACCACTGT	CTGCTCTCTA	CTTCCATGAC	ATATGCTTTG	TTTTAGCTCT	720
	CACATATGAG	TGAGAGCATG	CGACATTTAT	CTTTCTGGCC	CTGGCACATT	TTTGAATCAT	780
	TGTTAGAAAA	GATGATGGTT	TGGAGTAGAT	ACATCAGAAG	TGACAGCGTT	TGCCCTAAAA	840
	AGGAAAGACA	GGCTCCTCTG	GGACCCTGAC	CAAGTTCCTG	TGAACTATTT	TATTATTGTG	900
(CTGTGTTAGT	CCTGGGGTCT	TCCGTTCCCA	GCCCTCCTCA	CCTGCTCCCA	TATGGCTCTC	960
٠	CTCTTCTTC (CAACCTCTCA	GGATGTCCTA	TGAGGATTTC	ATCTACCATT	TCACAAAGTT	1020
(GGAGATCTGC A	AACCTCACGG	CCGATGCTCT	GCAGTCTGAC	AAGCTTCAGA	CCTGGACAGT	1080
(STCTGTGAAC (GAGGGCCGCT	GGGTACGGGG	ттсстстссс	GGAGGCTGCC	GCAACTTCCC	1140
A	AGGTGGGAGA	TGCTCTTGAT	GGGGGGAGGG	TCTAAGCCGA	AAAAGTTCCA	GGCAGAAGAA	1200

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GCCTAACTAG TGCTTATTAA GTCTCTCTGT TCCAGACGTC CACTATCTTA TTAAACCTTC 1260 CCTGTTTTAC TGAGAAGGAA ACCACCATGC TGAGAAGTTT GCAATAGGGA GCTGGGTAGC 1320 AACTTTGGAA GCAGGAACTT GTGGGAACAA TGCAGATGCT GCTTGGACTT ACGATGAGGT 1380 TATGTCCAGA TAAGCCCATC CATCTTTTGA AAATACCCTA AGTGAAAAGT GCATCCAATA 1440 TGCCTAACCC CCCAAACCTC ATAGCTTACC CTGGCCTACC CTCAAACATT GCTCGGAACC 1500 CTTGACCTTA AGCCTAAAGT TGGGCCAAAT CATCTAACTC CAAAGCCTAT TTTACAAAGA 1560 AAGTTGTTGT AATATCTCCA TGTAACTTAC TTAATACTTG TACCTAAAAA GTGAAAAACA 1620 AGAATGGTTG TACGGGTACT CGAAATCCAG TTTCTACTGA ATGTGCATCT CTTTCACATT 1680 GTAAAGTTAA AAAATTGTAG CCGAACCATC CTAAGTCAGG GACTGTGAGT ACTGTGTCAG 1740 TAACAGTAAG GGCACTATTG GAGAACCAAG TTAGCAGCTG CTGCAATAGT TCAAGTCAGA 1800 GATGATGAAA ACCTAGACCA AGTCAGTAGC AGCAGAGATG GAGGGGAGAC AGCAGATTTA 1860 GGGAGAGCAT ATTGGGTGAT GTAGGGAAGG AAGAAGAATG ATGTCAAGAT TCCCAGTTGG 1920 GGACCTGACA ACATTGCAAC ATAAGACACA CAAGAAGATC GGGTGGGTGG CTCATGCCTA 1980 TAATCCCAGC ACTTTGGGAG GCAGAGCCAG GAGGATCACT TGAGCCCAGG AGTTCAAGAC 2040 CAGCACAGGC AACATAGTGA CACCTCATCG TTACCCAAAA TAAAAAAAAA AATGAGGTGG 2100 GAGGATTGCT TGAGCTCGGG AGGTTGAGGC TACAATAAAC TGTGATCATG CCACTGCACT 2160 CCTGCCTGGG TGACAGAGTG AGACCCTGCC TCAAAAAAA AAGACACACA AGAGAAAAAT 2220 ATCAGCGTGT TGTTTGTTTT TGGTGGAGTT AATTGTGGGG TTCTAGGGAA AGGAATTTAG 2280 CTTGGGACAT GGAAAGTTTG AGGTTCCTGT AGAGTGTCCC AGTGAAGATT TGTAATAGAG 2340 2400 GGAATAAAGG AAAGAAGAGG CCAGACGTGG TGGCTTATGC CTGTAATCCC AGCACTTTGG 2460 GAGGCTGAGG CAGGCGGATC ACTTGTGGTC AGGAGTTCGA GACCAGCTTG GCTAACATGG 2520 TGAAAACCCA TCTCTACTAA AGATACAAAA ATTAACCGGG GATGATGGTG GGTGCCTGTA 2580 ATCCCAGCTA CTTGGGAGGC TCAGTCAGAA GAATCGCTTG AACCCAGGAG GCGGAGGCTG 2640 CAGTGAGCCG AGATCGCGCC ACTGCACTCT AGCCTGGGCA ACAGAGCCAG ACTCCGTCTC 2700 AAAAAAAAA AAGTGAGAGA GATTGAGGCT GGGATATATG GCTCAGGCAT CATGCGCGTG 2760 TAGGGGGCAG TTAAAAAGCA GAAGTAAGAA AGATTGCCTA GGGAGGCAGG AAGGGTGAGG 2820

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TGAGAGGAGA AGAGGCCCAG GACCAGATTC TAGTCACCAA CAGCGTTTAA GGGGCAGGTA	2880
AGGAAAACAA AACCATCAGC AAAGACTGAG AATGAAAGCC CAGAGAGGAA GGAAAAGCCA	2940
CACATACAAT CAGTACAGCT CCATCTGAAT AAAGGTAGCG CCCCCCCCC CCCAAATCAT	3000
TAGAGAAATG CCTGATTCGG TTTTCTGTGG ATTTTTCCTA AGAACCTAGA TGTGGGGAAT	3060
AGAAATAAAT GGTTCCCTCT GTCTCATCCC CTCCCTGCCC TCTGAGAGGA AGCTGTGATT	3120
GCGTGCTCCC TTTCTGGGGG TGCAGATACT TTCTGGACCA ACCCTCAGTA CCGTCCGAAG	3180
CTCCTGGAGG AGGACGATGA CCCTGATGAC TCGGAGGTGA TTTGCAGCTT CCTGGTGGCC	3240
CTGATGCAGA AGAACCGGCG GAAGGACCGG AAGCTAGGGG CCAGTCTCTT CACCATTGCC	3300
TTCGCCATCT ACGAGGTGTG TAGTCCTGAT TGGCTCCAGC CCAGGAAACA TACTTTCCCA	3360
GAGAGGACGC TTCCAGGGGC TTCTAGAGGG GCCCTCTGCT TCCTCAATAC CAGTGACCCA	3420
CAGAGCTCCT GGTATCAGGA CCACTTGTGT TTGTAACAAG CAAAAAATAC CAGGGGGGGC	3480
ATTAGAGAGG CAGTGGAGCG GGCCTGGCAG AACAGGTGCC TGGGGGTCAG GCTTCCGCAT	3540
GCGGGCTGCA GTTGCTGGCA TTGCCTTCCG CAGGCTCCTC ATCCTCATTC ACATCTGAAG	3600
CATCTTCCTT TCTGTTTCTT CTCAAGGTTC CCAAAGAGGT ATAGCAGCAG CAGCGGCCAG	3660
CAGTTGTGTG CAGCACTACC CAGGGGGGCC CGAGTCTGTC TGTGGCTCGT CGAGAAGCTT	3720
CCTGGTGGGG TTTGTGGGCA GGACTTGTGA TAGGAGAGGG CCTTGCCTGT TGTTATTTCC	3780
CACTTGCAGA GCAGGTTGCC TCAGGGCATT GCATGACCCA TGACTACCAC CCCCAGGATG	3840
TGCACTTTCT CCCTCGCACC AGACACTGCA CGTCACACAC ATGCCTTTGC ACACTCACCC	3900
TCCTCCACGC TTACAGCCAC ACACACAGTC ACACAGACGC GTTCTGAGGG TGGCTGCCCG	3960
CTTGGGATGG AGGAATCACT TCCCTCAGAA CCCAGCCAAG TCCTCTAGGC CTCCTTGGGG	4020
GTCCTTCCAG CCTGAGGGGC TTCGGAGCTG AGGACAGCTG TTCTGGTAAG TGTCCCTGAG	4080
TGTGGGGATG ACACATTTCC ATTCACTCTG AATCACAACA GAAAAGGGAA GAGGAATTGA	4140
GGTAGGGAGC CTATTTAACC CTTGGGAGTC GGGAAGTAGG GAGGTTGAAA CTGTGACATG	4200
GGTGACCAGG GAGTTGGGAA GGGACCCTTG GAGGTGGCTG TGGCAGGACA GGACGTTCCT	4260
CCCGAGGGGC TCATGTGCCC TGGGCTCTCC CCATCTCTCA GATGCACGGG AACAAGCAGC	4320
ACCTGCAGAA GGACTTCTTC CTGTACAACG CCTCCAAGGC CAGGAGCAAA ACCTACATCA	4380
ACATGCGGGA GGTGTCCCAG CGCTTCCGCC TGCCTCCCAG CGAGTACGTC ATCGTGCCCT	4440

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CCACCTACGA GCCCCACCAG GAGGGGGAAT TCATCCTCCG GGTCTTCTCT GAAAAGAGGA	4500
ACCTCTCTGA GTGAGTGCTG GCCCAGCTTT CCCACGTGTT TCTAAAAGCT CACATGGCCC	4560
ACTCCAGAGG TTGAAGGCAT GAGGCAGCTA GACACGTCTC CTCCAGGGTC CTTCTGCTGC	4620
TCCTGAGCCA CTGGCCACAT TACCCCCATT CATTCATTCA TCCATTCTGT GATATTTATT	4680
GAGCACCTAC TATGTTCCAG GCACTGTCCT AGGCACTAAG GATAGAGTAG TGAAGTAAAC	4740
AGAAAGAAAT CCCTGCCTTC ATGGAGCTTA ATATTCTAAC ATGAGACAAT AATGGATAGG	4800
AAAAACATAT GTAGCATGTT AGATTTGGAG AGGTGATATG GAGCAAAAAT AAAGTAGGGA	4860
AGAGGGATAG GAGGTGTTGG GGATGCTTGA AATTTTAGGT TAGCATGGCC AGGAAAGCCA	4920
CATCCTGTCC CTGGCCACCA CAGATGAGCT CATAGCCCCT GCCACTCTGA TCTCTGTCCT	4980
TGGAAGATGC ACCAGGTCCA TGGGTAGGTG GCTGGGTCAT GCCTTTGGGG GGCTCTGAGC	5040
AATACTAACA AGAACCTGCG TGCCTGGGCT TGGCTGTCGG GGATGGTGCT GACATGGGGC	5100
TGGTTCCTGG GGTTGGGGTG TTCCAGGGGT TCTCTAGAGG CTGGTTCTGG CTTGGCTGCC	5160
AGGAAGCCGT GCACCAGAGC AAACCGTCCA CGGGCCTCCT GCTTGCTTCT GGTGACACTG	5220
AGACCCCACA TGTCTGTATT CCTCACAGGG AAGTTGAAAA TACCATCTCC GTGGATCGGC	5280
CAGTGGTGAG TGGTTTAGAT CTTCTGTGCG AAAAGTCCAG AGGGTCCCCT TCCCTGACCA	5340
TGCAGGGGAC AGATGGTGCA GGGGAGAATG GGCACTGGCA GAGGGAATGG GAGTCTGGGC	5400
TGTGCTGAGC AGTCCCTCCT TGGCACTGCA AATCCTACTT TGGCATGGCC AGAAGTAATC	5460
GGCCTTAAGC ACCGGGGCC ATTGAGGCAG TTCAGGGGCT GGGAAATATG GAAGAGGGTC	5520
CTGGAAAGGA GAAGCAATTT GAACAATCGG AGGGAACAAG GCCACAGGAA GGGATGACAA	5580
GAGCCGCAGC GAACACTGGA TTCTGAGACT GGATAACATT GGATTTCACA CATAGAGAAA	5640
AGAAAGTAAG CTGGTGCCGG ACCTGGTGTT GACACTTGGA TCCTCCACTT ACCAGCGGGG	5700
TGACCTGGAC AATTTCTGTA ATCCCTCTCA CTCAGTTTCC TACTCAGTAA AACGGGGATG	5760
ATAATGTGCC TTGCAAGGCT TTTGTGAGGC TTCATCAATG AGGTGATGTA TGTGAAGTGT	5820
CTGGCACAGC ATGGGCACTC AAACAGAGGT GCTTTTCAC ACTTTACACC TTACAAGGTA	5880
CTTTTCACAT GTGTCATCGC GATACTTGCA AGGTTGCTGA GAGGTAGATG GGGTTATAAT	5940
CCCTGGTGTT CAAGAAAGGA AGCAGAGGCT CAATGGGGTT GAATGACTTC TCTGAGTTCA	6000
CAGAGCTCAG TAAGTGGCAG GGTTTGGAAC TCACATTCAG ACTCTCTGAC TCCAGACTTA	6060

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GGTTTTTCCG CACCTCCACG CTGAGGCCAG CCCCAGGCAG TGAGAAGCCC AAAGTCCGAA	6120
GCACAGAGTG CTGTGTTG GGCTCTGTGT GTTGAGGAGT CTTGTGACTG CCTTGGGGCT	6180
TTGGGCTGTA GTCAGCTGAC AGTCCTTTGT GCTCTGTGGG GATGACGTAG GCCAATGGGA	6240
GGACAAATGC CCCTCTGAAC TGTCTTCTGG GCAGTGACAG TCATGGTCAT AATCCTGACC	6300
CTGAGCCAGT GCCAGGTCTC CAAGTGCCTT CTGAATGACC ACAGGCGATT GGTTTTAGTG	6360
GTAGGTGCGT GGGGATCTGT TCTGGTCATC TGGATGCTGG TCATCGGGTG CAGTATTGAT	6420
CAGGACCTGC AAACCCAAAA GCTTATGGGA GCTGGCACGT CACGTGAGTA GAGCAGGCAG	6480
GTGCAGGGTT TTTGATGTCC CTGCACTGAC ACAGTTGTCT GCAGTTCTCC AATTTGACAT	6540
TTGGGCTCCA GTGTCGAGGG TCAAACAAGG AATTTTGGGG CGTGGGCCAA ATCTGGGAAG	6600
ACACAGGGAG CAGGGCCCTT TGGCTCAAGC TGATAGTTGC CGCAGGGATT ACCAGGCCCA	6660
GGGCAGCCTG CCACAAGCTG GGGCTTTTAC CAAAGAAAAT CTCCCTATGT TAAATGCTTG	6720
CTCAAAAATT TTTAAAAAAT ATTCTGTAAG TCAAAATCCA TTGTTAGGTC AGTTTGAGAG	6780
AGCCATGTTT TTGGTGTTTT AGTAACCAAT TTCATTTTTT TATTATTTAT TTATTTGTTT	6840
ATTTTTGAGA CGGAGTTTCA CTCTTGTCAC CCAGGCTGGA GTGCAATGGC ATGATCTCAG	6900
CTCACTGCAA CCTCCGCCTC CCGGGTTCAA GCAATTCTCC TGCCTCAGCC TCCTGAGTAG	6960
CTGAGATTAC AGGTGCCCAC CATCACGCCT GGATAATTTT TGTATTTTTT AGTCGAGATG	7020
GGGTTTCACC ATGTTGGCCA GGATAGTCCT GAACTACTGA CCTCAGATAA TCCGCCCACC	7080
TCAGCCTCCC AAAGTGCTGG GATTACAGGC ATGAGCCAGC ACGCCCGGCC ACCAATTTCA	7140
TTTTTTAAAA AAGGAAGAAA GAAAACCTTA GCCAGAAGAT CTTTTTCCTT GCCATATGCA	7200
GTAAGAGTAG ATTATAAAAA CAAAGTCAGA GCAGTCACTG GTGTCTGGGC ATGGAGGAGA	7260
AAGAAGAATT CTCTTCTCCC TTCACCCTCC ATGCCCCTTT TTGGCTCCAT GTGATTCAGA	7320
TTTCTGGACC CTGGAGCCCC ACCCCAAGCT AAAGACCAGG ATACAGGGAA GCCACAACCA	7380
CTGGCGGTTC TGAGAACTTA CTTTTCACTT ATTCTGCATT TACTGTTTCC TTTTCTTATG	7440
CAGAAAAAGA AAAAAACCAA GGTAGGTGTG TGGGTAGAGA GCATGAAGTG TGTGTACTCA	7500
TGCATATGTA TGTGCATGCA TGTGAAGTGT GCATGTGTGA GCTCATATGC ATCCATGCAC	7560
CAGACTTGCC TCTTCCTCCC CCTCCTTCCT GAGCTTCTGC TGGGGCCGAG CGTGCAGTAA	7620
TGACAACTAC GATTTGCTGG GGGAAGGCTA CGTGCCAAGC ACTCTTTTAG GTGCTTTCCA	7680

FIG. 8D/5
SUBSTITUTE SHEET (RULE 26)

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TGATTAATTC CTTCCTCACA ACACCCCT	
TGATTAATTC CTTCCTCACA ACAGCCCTAT GAGATTAGTA CTATAACTAT CCCCATTTTC	7740
AGAGGGAGAA AAGGTACAGA CTTGACTAAC TTGCCCAAGG CCACACAGCC AGAGAGGGGG	7800
AGAGCCAGTA CTTAGAGCCA GGCAGTCTGG GTCCAGAGTC CGTGTCCTGA ACCACAAGAG	7860
GCCATCATAC GCCATCAGAT TTGGTGCTAG CATTTCTGGT GGTGCCTGGT GGTGATGGAT	7920
CCATCACAGG GGTCCTCCAG GTACTGGTGC TGGCCCAGAC CAGAGCTGAC ACTCCTCAGG	7000
CACTACCACA TTCCAGGCAC TGTGCTTGGG GTCAGTCCCT CTCTTTTTT TCCCCCCCAA	8040
TTATAACAGT ATCTACAAAG TAGGTGCTGT TATTTTTCCC CTTTCACAGG TGAGATAGAC	8100
TCAAAGAAGT GAACTTGCCC AAGGAACAGA ACTAATGAGT GGGGAAAATG GAACTGGAAA	8100
CCATGTCTGT TTACTCCAAA ACCTGTGTTT CTTGCCCTCT TTCTCTGATG CCAGCCCCCT	
ACACTTCAAG GCCTGTGTTG TCCAGACCCA CACTCGGGCC TGCCAGTGTG TGCCTGGCAG	8220
GGATGCTCCA TGGCCACACC ATATCCATCC TACACATCCC CCCTCAGACT GTGACCTCCA	8280
TTTGCTCTGG GATCCCGACA ACCTTCAGE	8340
TTTGCTCTGG GATCCCCACA AGCTTCAGCT GCTTGAGCAA GACACTGCTT AGAAGGCAGA	8400
GCAAGCCAAG GCCTCTGGGG CCTGCTGGGA GCCAAAGCTG GGGAGCCGTT TCCACGGGTC	8460
TATCTGCTTG AGCTGTCCTA GATGAGCAGC ATGGAAGGCC AGTGGTGCAT GAGTCCAGGC	8520
GGGCTGCTTT TCTGCTCCGA GAGGCTCTGC CTGCCCAGTT GTTCTCTGCA TTGCAGCCTC	8580
AATCCCCACA GCCTTGCCTT CCCCCGGCTT TCCCTACAGG TGCACCGCAT CCACAGTGTT	8640
GGCACCATGC AGCAGCCGCT CTCCGTCCTT TTCATATCCT TGTCACTTGC ACGAGCATGT	8700
CTTGAAAATA TCCCTTGTTT GTGTAGCATC TTAAATGTTT TTGCAGTATG ATTTTGCATT	8760
CAGTATCTCA TTTGATCCCC ACAAGAGCCC TATGAGGAGG GAAAGCAGAT TTTACCATTA	8820
AAGGATGAGT AAACTGAGGC CAGAGAGGAT ATTTTTGGTT TTTTTTGAGA CAGTCTCACT	
CTGTCACCCA GCCTGGAGTG CAGTGGCTTG ATCTTGGCTC ACTGCAAGCT CCACCTCCCA	8880
TGTTCACACC ATTTTCCTGC CTCAGCCTCC CAAGTAGCTG GGACTACAGG CACCCACCAC	8940
CACACCCAGC TAATTTTTTT GTATCTTTAG TAGAGATGGG GTTTCACCCA GTTAGCCAGG	9000
ATGGTCTTGA TCTCCTGACC TTGTGATCTG CCTGCTTCGG CCTCCTAAAG TGCTGGGATT	9060
ACAGGCGTGA ACCCCCCTGC CCGCCACAC ACCATAGTTG	9120
ACAGGCGTGA ACCCCCCTGC CCGGCCAGAG AGGATATTTC TTAATGAGGG GCAGGGCTGG GATTCCAGCC CAGTGTTCTC ATCCCTGAGG	9180
GATTCCAGCC CAGTGTTCTG ATGGCTCACC CACTGACCAT TCCACTAATC CGTGTCCTTT	9240
TTCAATCTAA ACTTTCAGGG TTGTAGAGGT TCCTTTGAGG TGCCTCAGTA CTTCCATGGT	9300

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0.50	
GATGTGGGGT CTGAGGGCCA AGAGCTCTGT TCTCATTAAT CAGAGAAGCT TGTGTTTTTA	
AAAACACCAT GTTTACTGCA GGAAATTTAA TTGGACAGTG TTTCCATCTG GAAAAAAAA	
AGTCTACAAA ATACTTGACA ATCACTGCAC TAGATCATGC TGCTTTTAGC ATTCTTAGCA	
TTTCACGTGC TGAGCTCTCA ATACTCTACC ATGAGGAGGG ATGGAGTGGG TATGAAAAGA	
TAAAGAACTG AAGTCACACG GCTTGTCAGT GGCAGAGATA GAGCTTGAAC CGAGGTTGAA	
GAGCTCCCGC CTATTCCTTT CCTCTTCTCA CTGGATAAAG CTGCTCCAAG AGAGGTGCTG	
CCTCAGTGTG CCTGTTCAGA CTGTAATCCT CCCTTCCTTC CTGCCTCCTC CCTCCTCTCT	
CCAGCCCATC ATCTTCGTTT CGGACAGAGC AAACAGCAAC AAGGAGCTGG GTGTGGACCA	9780
GGAGTCAGAG GAGGGCAAAG GCAAAACAAG CCCTGATAAG CAAAAGCAGT CCCCACAGGT	9840
GTCTGGGCAT GTGGCATGGG TGGGGTGGCC AGCACGCTAC AGGGGCTTCC TATGCGCTTG	9900
GGATACACAG GGGCTGGAGG CTTCCCAGGA GTTTGTCTTG AACATCTGGA GGTTTGAATT	9960
TGTCCCACTG ACCTTTCTT TCAGCAAGTT CCCCTGAAAT TTGGGCTGCT GCTTGGGTGA	10020
ATATCCCAGG ATGGGGGTTC CATTCTAGGA GTGGACTGGC AGGCTGAGCC TCCCATGGAG	10080
CTGATCCAGC CAGGATACAG AGAAGGGGAG GCAAAGGCTG AGACAGAACC AGCTTGAGAG	10140
CGGAGGCGCA ACTCTTGTCT CCTGGTGGCC TTGAGCATTT CACAATAGGG GGATAAAGGA	10200
TAGGAGCAGA AAAGTGGGGC TGACTTCAGA AATGGGGTCC TCTAGAGCTC ACGGGAGGGT	10260
GTTAGATTGG AGTGGGAGCT TAGTGGAGGT GAGCCTTAGA GGCAAAAGTC TCCAGACCAA	10320
TCCAGGCCCC CTCTTCTATC CGGGGGCCCCC TCTTCTATCC AGGGCCCCTC TTCTGTCTGG	10380
GAGCCCCTCT TCTATCTGGG GCCTCATGCA GTGGGGCCTA GGGGAGGTTC TCTGAGGACT	10440
TGGCCTTGAT GACAGGGTGG CTGGAGGAAT CAGAACGGTC AGACCTTCTT TGACCTGCGG	10500
GCACCTTTAG TTGGAATGCT CAGGCCTGGG ATGGTGGAGG GGGCTCTTGC AGGTGGGGAC	10560
TGGGGTGGCG GGGAGGAGGC TGTATGGCCG CCATATCTCC TTTGGCTGGG GGCGTCAGGG	10620
CTGGAGAGGT GTGAAGAGTC CCTGAGGCCT CGATGCATCT CACTCCAGCT CACCAGGTCT	10680
GCATTTGCCC GTCCCCAGCT CCTGCTGCCA CCCCCGGCCG TTTTAGGCAC TTGGCTCCCT	10740
TGGCCCAGAG GAGCTTGCCT CACAGGCCTG TGCACCTCTG ACCCCTGTGA ACCAGTTTTC	10800
CTTTGTGCCT CCACAGCCAC AGCCTGGCAA CTCTGATCAG GAAAGTGAGG AACAGCAACA	10860
ATTCCGGAAC ATTTTCAAGC AGATAGCAGG AGATGTGAGT ACCTCCAAGC CCAGGACGCC	10920
	-0720

FIG. 8D/7
SUBSTITUTE SHEET (RULE 26)

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CACAGGTGCT TCCTTCTCTC CTGGATTAAC TGCTCAGATT ACCAATTATT TCATTATTG	
TTGGTAGAGG TCACTTTCCA CTTGCCT	T 10980
TTGGTAGAGG TCACTTTGGA CTTCGGTGGA GCCAGGGGAT GTGTGCGTAG CACACAAAT	11040
CACAAGCCCT TGAGTTTTGG ACTGCCACGT CTGCTGGGGG GCTCAGAGGC CTTTTTGCTC	11100
TGAGCTGCCC ACGGTGGTCC TGATAGCTGA GGTGCAGTAT CTGGCCCCCT GTCTTCCTCA	11160
GAAAAGCCCC AGCTTCCCAT GACATAATAG CACCGACAGG GATTTTACAA ACACAGCCAG	1100
GTGGAATTTG TTTTGCAAAG TGTCCGCGCC AGGAGCTGCT GTACTCCTGA ACCATGACCC	11220
TCCTCTCCCT TCCTCCTCAG GACATGGAGA TCTGTGCAGA TGAGCTCAAG AAGGTCCTTA	11280
ACACAGTCGT GAACAAACCT GAGTTOOTTA	11340
ACACAGTCGT GAACAAACGT GAGTTGCTCA AACCAAATGG GGGTGGGGTG	11400
CCCGTTGTCT CAAAGCAGCT CCTCACTCTT CTCCATCCCC CCAGACAAGG ACCTGAAGAC	11460
ACACGGGTTC ACACTGGAGT CCTGCCGTAG CATGATTGCG CTCATGGATG TATCCTTCCT	11520
GCCGCCCTT CCCGACCCTC TGTCATCAGC CCACGGGGGC CAAGGCAACA TACAGGGTGC	11580
CCAGTCAGGC AAAGGGCCCT AATTTGTGCC CAGGGAAACT TAAGGAGACC CTGATTCAGA	
ACATCTTGGA TACTCGTCTG AAAGGGGTTG TTAGAGGCGG AAGGGGAGGA TGTTGGGTTG	11640
TAACTGCCCT AACCCCTGTG CTTCTCTCAG GCCTGGGATC CTGCCCAAGC AAAAGTGGTC	11700
CTTAGGAGAG CGCCTCCTCC CTTAGAGAGT GCCTGGGATC CTGCCCAAGC AAAAGTGGTC	11760
CTTAGGAGAG CGGCTCCTGG GTTACAGAGT AGGCGCAATC TCTGACTGGT GGTGGAGTGG	11820
AGGGGAGGT TAAATAGTAC AACAGGGCAG TGGGTAGGAC AGCCCGGAGT CTCCTAGACC	11880
CTCCCTCCAA ATCCAGGGGG ATTTTGCTGT GTGCTGTGTA GCCCTGACCT CCCTCCTCCA	11940
GACAGATGGC TCTGGAAAGC TCAACCTGCA GGAGTTCCAC CACCTCTGGA ACAAGATTAA	12000
GGCCTGGCAG GTGGGAAGAG AAAATGAAGC GTGGGAGTCA AGAATGGGGT TGATTTGGAG	
ATTCAGTGTG TGACCTCCAT CCTCAAATTT TCTATTGCCA GAAAATTTTC AAACACTATG	12060
ACACAGACCA GTCCGGCACC ATCAACAGCT ACGAGATGCG AAATGCAGTC AACGACGCAG	12120
GTGCTGAGAA GGAAGGGGTC TCAGGGATTOT COLORS AAATGCAGTC AACGACGCAG	12180
GTGCTGAGAA GGAAGGGGTG TCAGGGATGT GGACCCGAGA CGGTGGGAGC AGGAATGGGA	12240
GGGGACTAGC TACTAGGGCC CCACTAGAGA AGGAGAGGGA AAGGGCTTCT CACTTTCCCT	12300
TCCCAGGTCA CAGAGTGTCC GAGAGGCAGG GAAAATAGAA GACAGGCCCA AGGCCTCCAG	12360
CTCCACGTCC ACCTCTAACA TGGTCCCCTC CACAGGATTC CACCTCAACA ACCAGCTCTA	12420
TGACATCATT ACCATGCGGT ACGCAGACAA ACACATGAAC ATCGACTTTG ACAGTTTCAT	12480
CTGCTGCTTC GTTAGGCTGG AGGGCATGTT CAGTAAGTGG GAGAGGGGGG CTGCCCTCTG	
The control of the co	12540

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CTCTCTTCTCC	
CTCTCTTGCA GGGGCAGTTG TGGCAACAGG CATCTCACCT GATAATCTCC AGTCTGCTCG	C 12600
ATCCAGGCTG AACAAGGGCC AATGACCTCT TTAGGCCCAG AATGGGATGG CAAAGGGAGG	3 12660
GTTACTGGTG ATTCTCTGCC TGCACATCTT TGTGCTGATG AGGGACAGCA CTGGGCACAC	12720
GGTCCTCTGA GGGGAAGTTA CAGTAGTAGA GGCGGAGTGC GCCTGTAACT GGCCTCTGGC	12780
CTGTGCATTC TTTCACAGGA GCTTCTCATG CATTTGACAA GGATGGAGAT GGTATCATCA	12840
AGCTCAACGT TCTGGAGGTA AAGCATAGGC ACAGCACATT CCCCCTACAC ATTAAAACTC	12000
AAGGTGGAGG GGTCAACGGG GCGGACTGGA CCCAGGGTGT GCTCCTCATT TCCACACAGT	12060
GGTGGAGGGA AGGGATAGGA ACAGAACATG GAGGGAGGCT CAGCAGGCTC CCAGGACACA	13020
TGCACTTGAG GCCCAAAAGG ACCTCTGCTC CCCCAGTCAC TTGATGCGGG AAAACATGCA	13080
CCTTCTTAGG GAAGATCTAG GAGAAAGGAA ACAGTAAGCC ACTGCTTCTT GGAAAATCTT	13140
CTGGGGGTCT GACCTGCTGG GACTGTTCCC TTTCCTCTTG CCCCGTAAGA TTCCTAGGGC	_
GGGGGGGGG GGGGTCACT CTTTTCTGAT CTACATTCTG ATCTTGGGAC TTCTTTCAGT	13200
GGCTGCAGCT CACCATGTAT GCCTGAACCA GGCTGGCCTC ATCCAAAGCC ATGCAGGATC	13260
ACTCAGGATT TCAGTTTCAC CCTCTATTTC CAAAGCCATT TACCTCAAAG GACCCAGCAG	13320
CTACACCCCT ACAGGCTTCC AGGCACCTCA TCAGTCATGT TCCTCCTCCA TTTTACCCCC	13380
TACCCATCCT TGATCGCTCA TGCCTAGCCT GACCCTTTAG TAAAGCAATG AGGTAGGAAG	13440
AACAAACCCT TGTCCCTTTG CCATGTGGAG GAAAGTGCCT GCCTCTGGTC CGAGCCGCCT	13500
CGGTTCTGAA GCGAGTGCTC CTGCTTACCT TGCTCTAGGC TGTCTGCAGA AGCACCTGCC	13560
GGTGGCACTC AGCACCTCCT TGTGCTAGAG CCCTCCATCA CCTTCACGCT GTCCCACCAT	13620
GGGCCAGGAA CCAAACCAGC ACTGGGTTCT ACTGCTGTGG GGTAAACTAA CTCAGTGGAA	13680
TAGGGCTGGT TACTTTGGGC TGTCCAACTC ATAAGTTTGG CTGCATTTTG AAAAAAGCTG	13740
ATCTAAATAA AGGCATGTGT ATGGCTGGTC CCCTTGTGTT TTGTTGTCTC ACATTTAGAT	13800
ATCAGCCATG CATGACTGAA TGGCTTCCAA TCATATACTC ACCTATCACC TACAAGAGAA	13860
CAATGAAAAA CACACAAAA AACAAAATCT TGAATTTTGT AATCATGCCT ATTGCTATTT	13920
CTTGAGCATA AGAATGGCTC AGATACTTTC CAAGACATAA AAGGAAGGCA GAGGAATAGT	13980
TGTTGCTGTA AAAGACATCA AGAATAAATG GGGTCATGTA CAACGGGAGG GGCCGGTTAC	14040
CTGAATAATG GAGTGGAGAT TGAGCTATCC TAGGTGTA CAACGGGAGG GGCCGGTTAC	14100
CTGAATAATG GAGTGGAGAT TGAGCTATCC TAGCTCCTCT GCTCACTAAC TGACCTGTCG	14160

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					CTGGACCATG	14220
GCCTGCGGCA	TATCTATAGG	CATCCTGTGT	TTTCCACCCA	GTTTCCTTCT	TCCTCGCTAA	14280
GCCAACGTGG	AAAGGGCTGG	CCGTGAATAT	GCAGACAAGG	TAACGAAAGT	AAACCGTCAA	14340
TTAGTAAAAG	TACTTCATTT	TCCTCTTGTA	TTTGCTTCAT	TCTTGCTTCA	CAAAGTTACG	14400
AAGTCCACAG	CTTTATACCA	AAATGTAAGA	AGGCTATTTG	CTTATAAACA	TTTTGAGTCA	14460
GGTGTCATCT	GATTTCATTC	TTCTAATCCA	TATTCAATAT	TAAAAAATCA	GAAACCAAGG	14520
		CATATATTTC				14580
		TTTCCCAATT				14640
	ATGTAGTCAC					14040
						14664

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Date of mailing (day/month/year) 23 July 1996 (23.07.96)	in its capacity as elected Office
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International filing date (day/month/year) 21 November 1995 (21.11.95)	Priority date (day/month/year) 22 November 1994 (22.11.94)
Applicant	
BECKMANN, Jacques et al	

The designated Office is hereby notified of its election made:	
	X in the demand filed with the International Preliminary Examining Authority on:
	19 June 1996 (19.06.96)
	in a notice effecting later election filed with the International Bureau on:
2.	. The election X was was not
	made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland Authorized officer

Mirjam Van Straten

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Facsimile No.: (41-22) 740.14.35



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(PCT Article 20)

Date of mailing:

10 October 1996 (10.10.96)

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International application no.:

PCT/EP95/04575

International publication no.:

WO96/16175

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